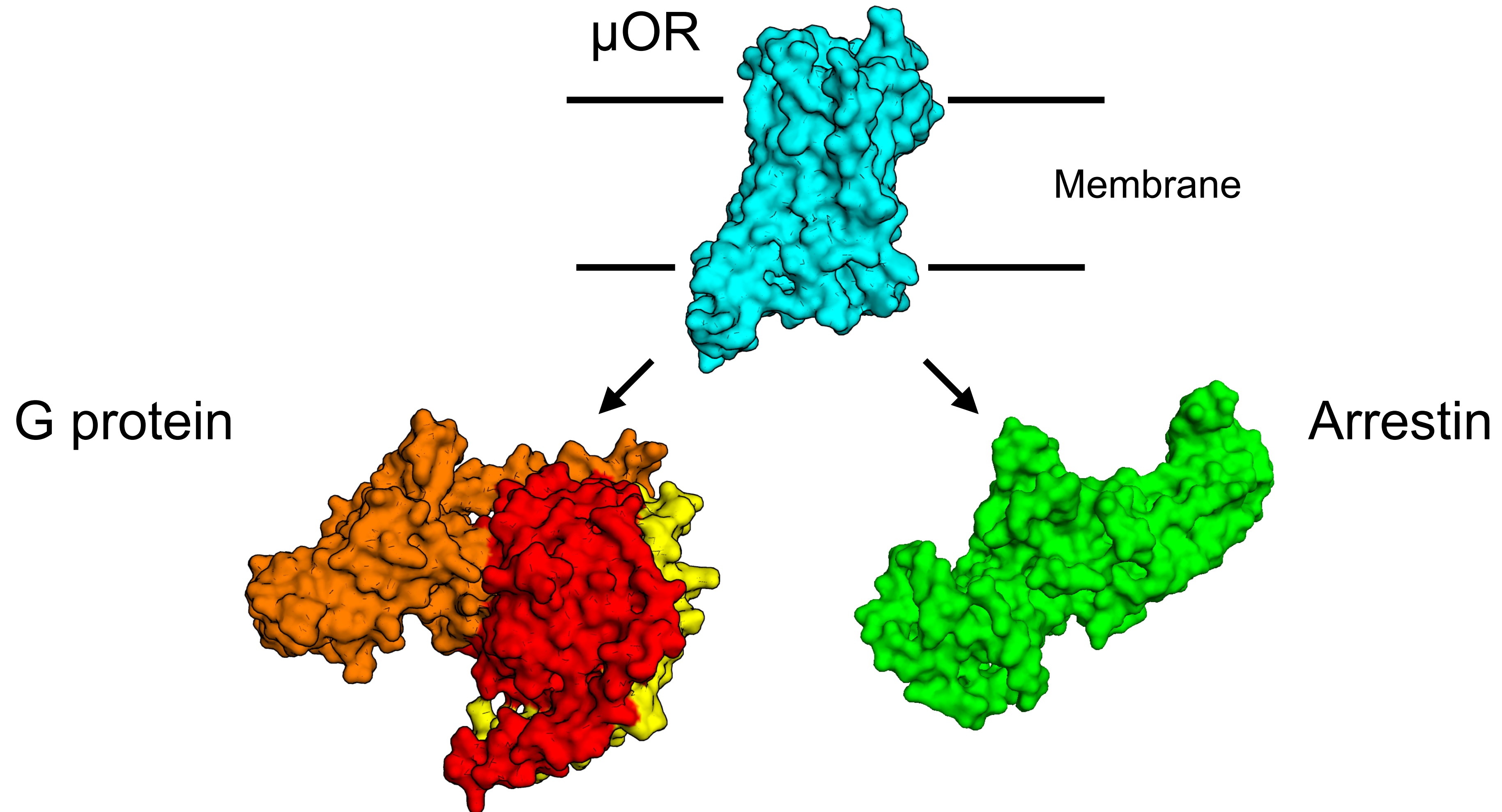


# How ligands achieve biased signaling at opioid receptors

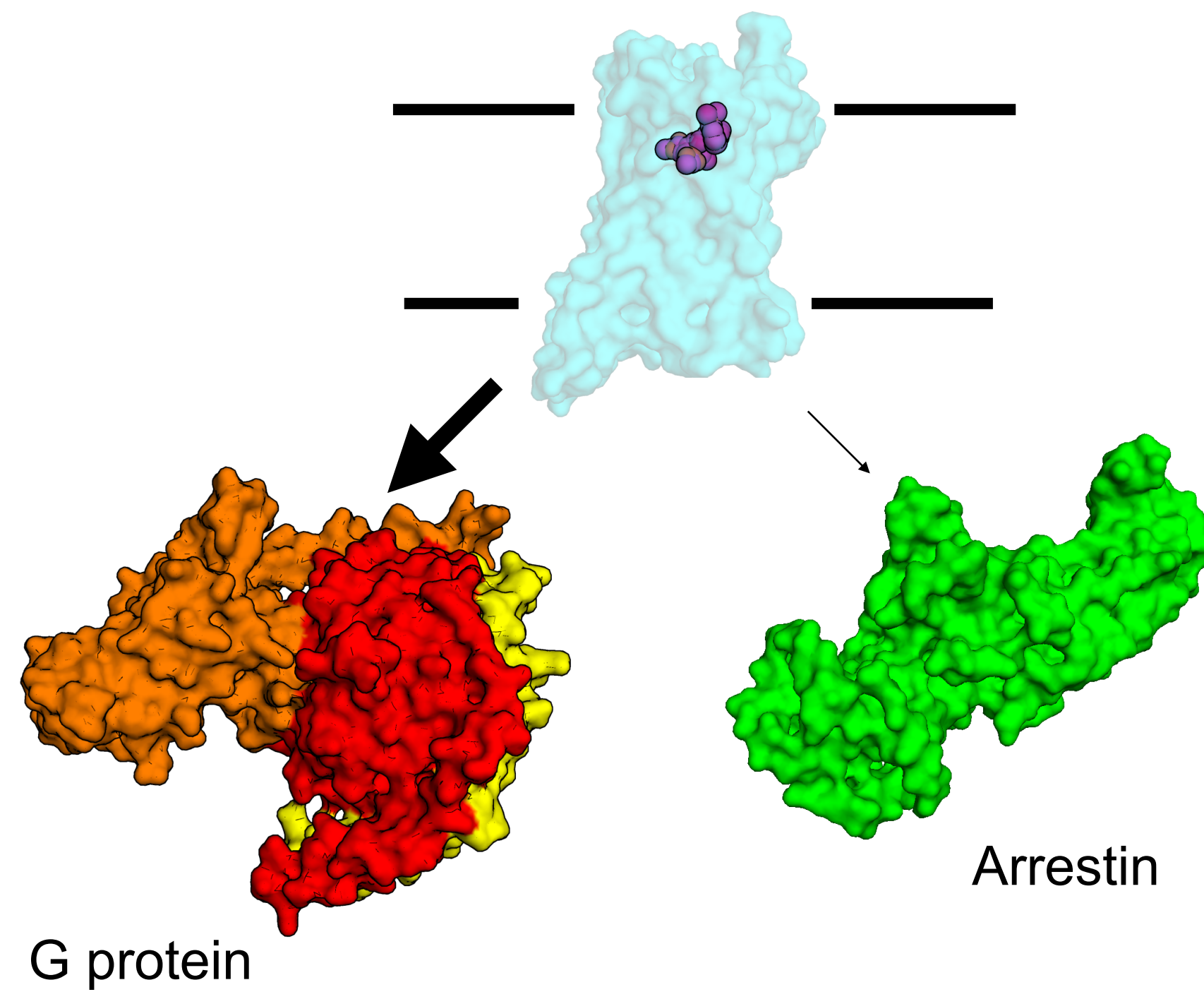
**Joe Paggi, Ron Dror lab, Stanford University**

# G protein coupled receptors (GPCRs) signal through multiple transducers

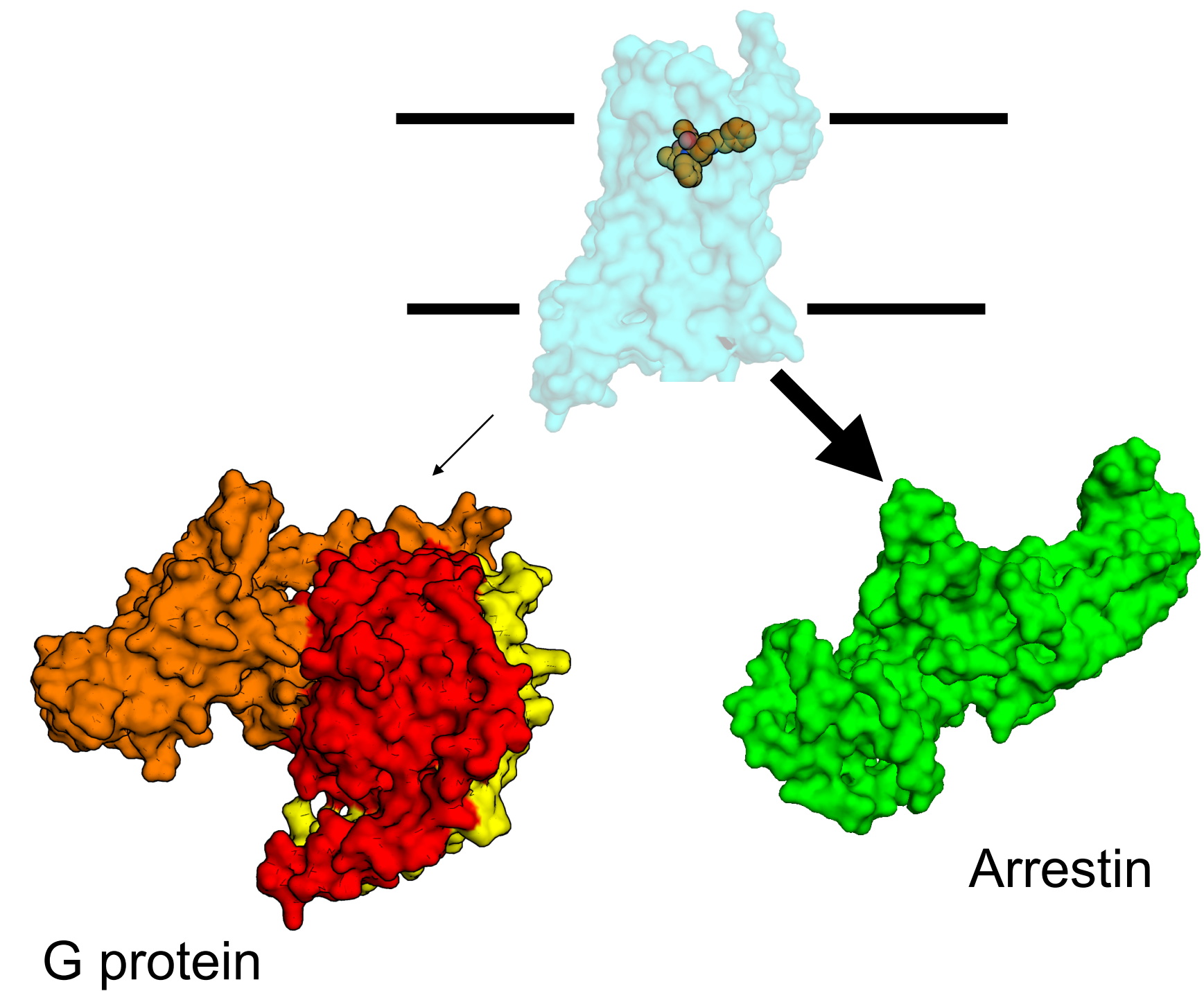


# Biased signaling

G protein biased agonist

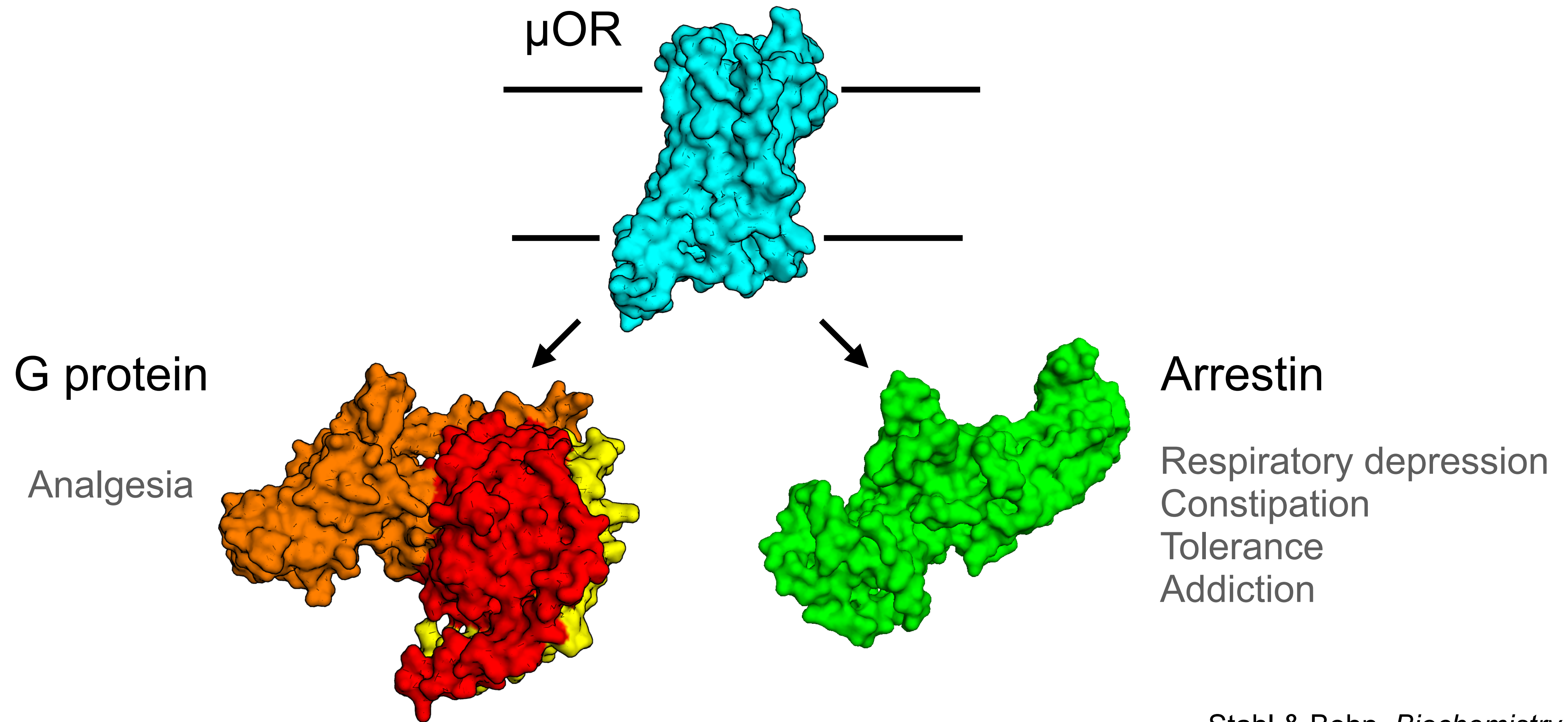


Arrestin biased agonist



Always **relative** to a reference agonist, I'll call these "balanced"

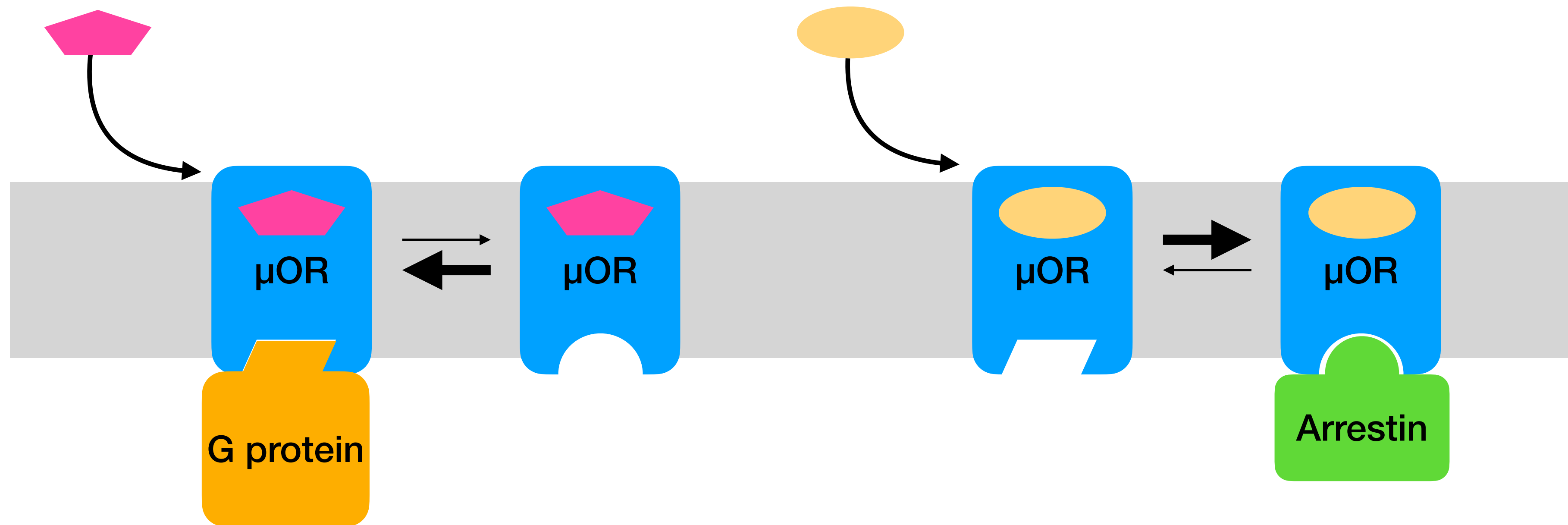
# Biased signaling promises safer drugs



Stahl & Bohn, *Biochemistry*, 2022

There is ongoing debate about the importance of bias signaling, as opposed to partial agonism.

# Biased agonists favor “biased” receptor conformations



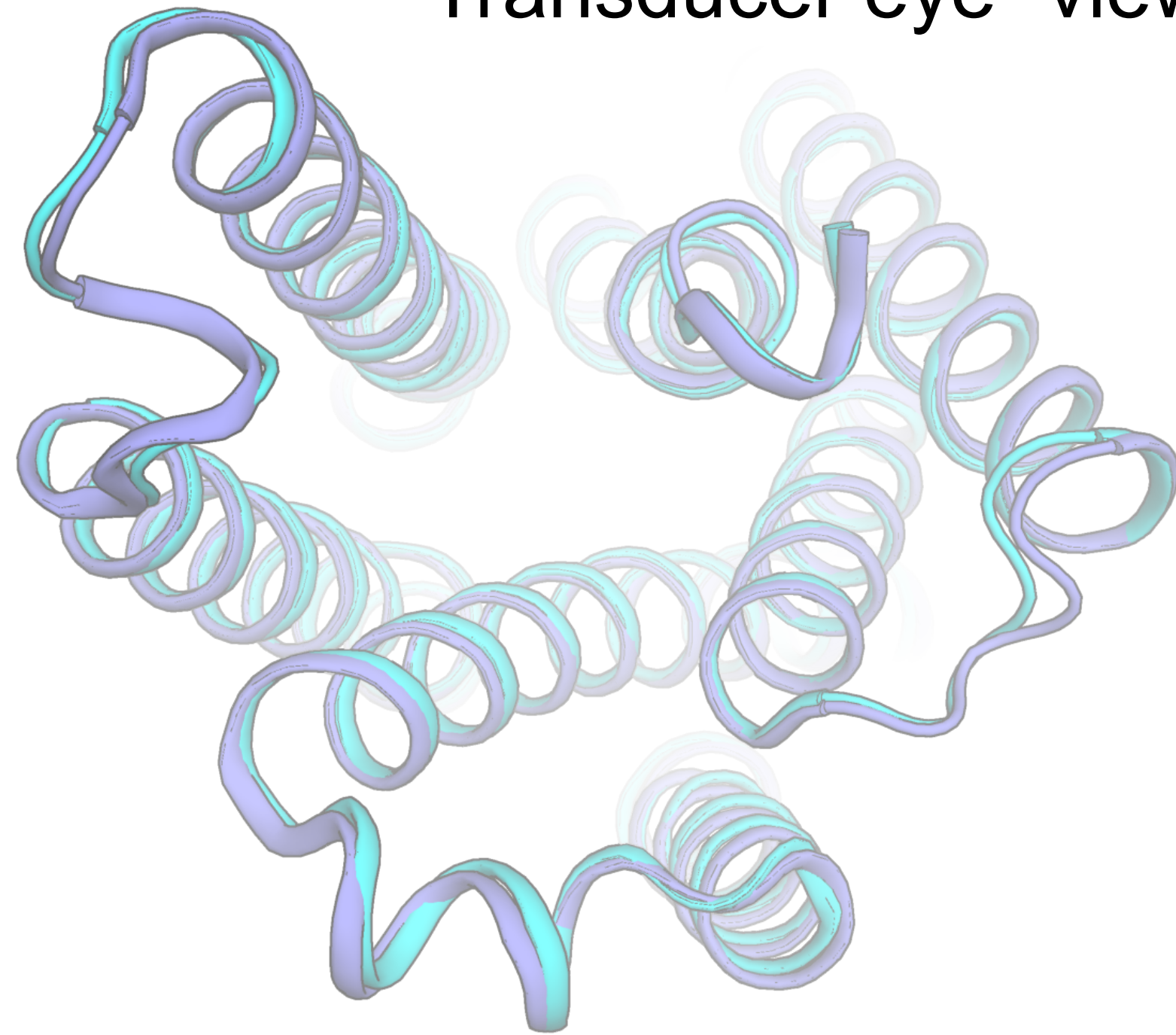
A variety of biophysical experiments support that biased agonists favor distinct receptor conformations.

- Liu et al, *Science*, 2012. Wingler et al, *Cell*, 2019. Cong et al, *Molecular Cell*, 2021.

- 1. What are these “biased” conformations?**
2. How do agonists favor different conformations?

# Experimentally determined structures provide an incomplete picture

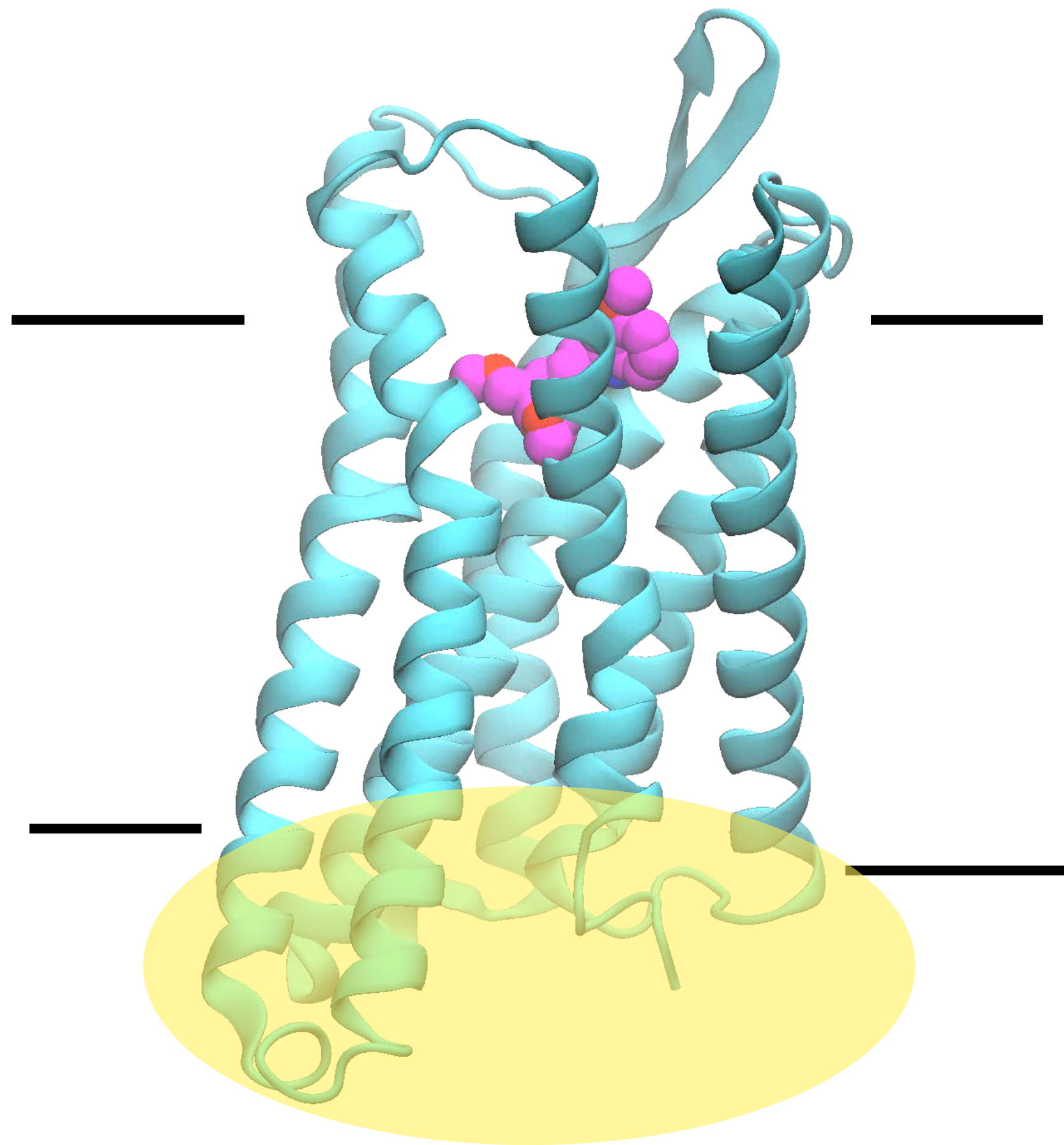
“Transducer eye” view



$\mu$ OR bound to arrestin-biased ligand  
 $\mu$ OR bound to G protein-biased ligand

- Solving an active state structure **requires** a transducer to be bound
- The conformation of the transducer binding site is largely determined by the transducer

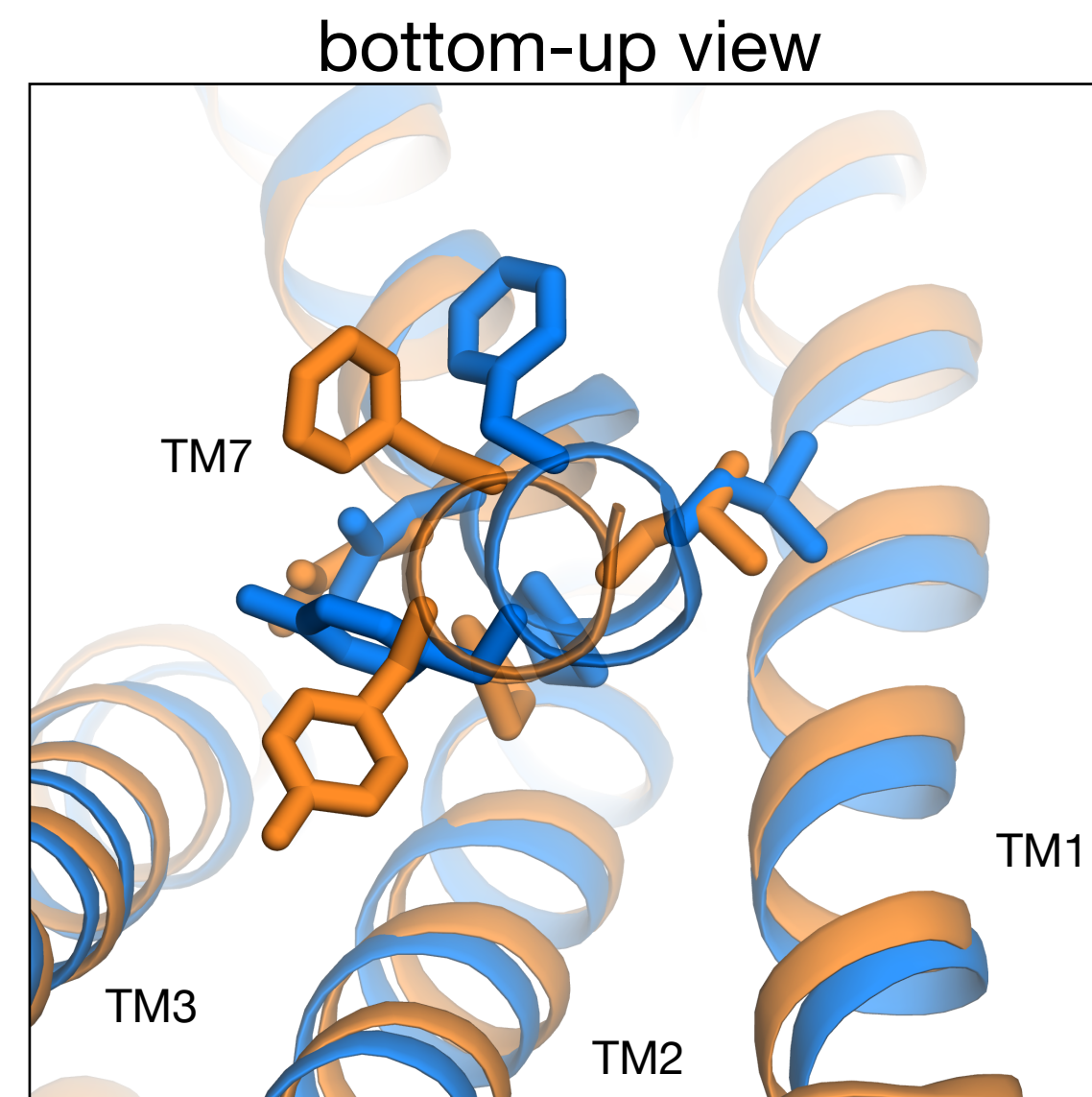
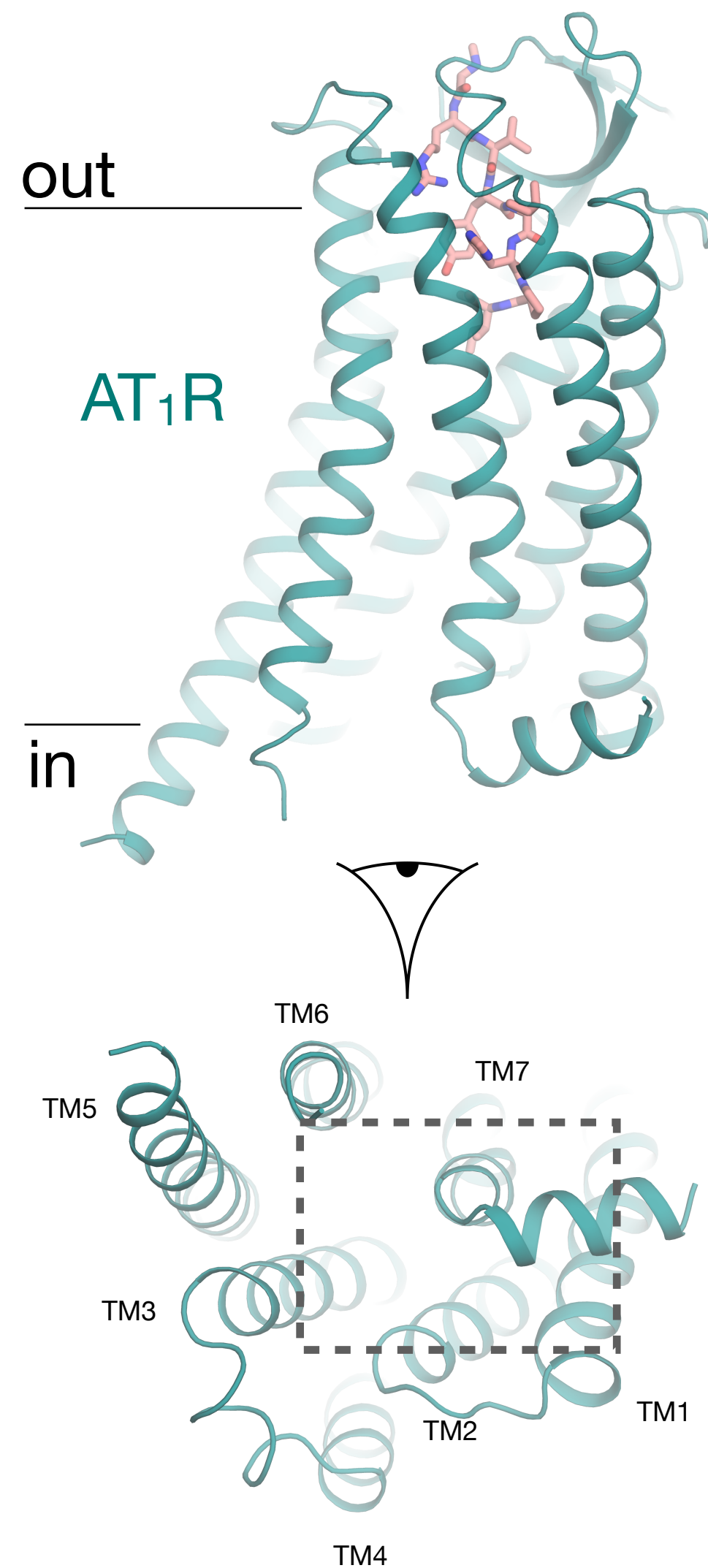
# How we study biased signaling using MD simulations



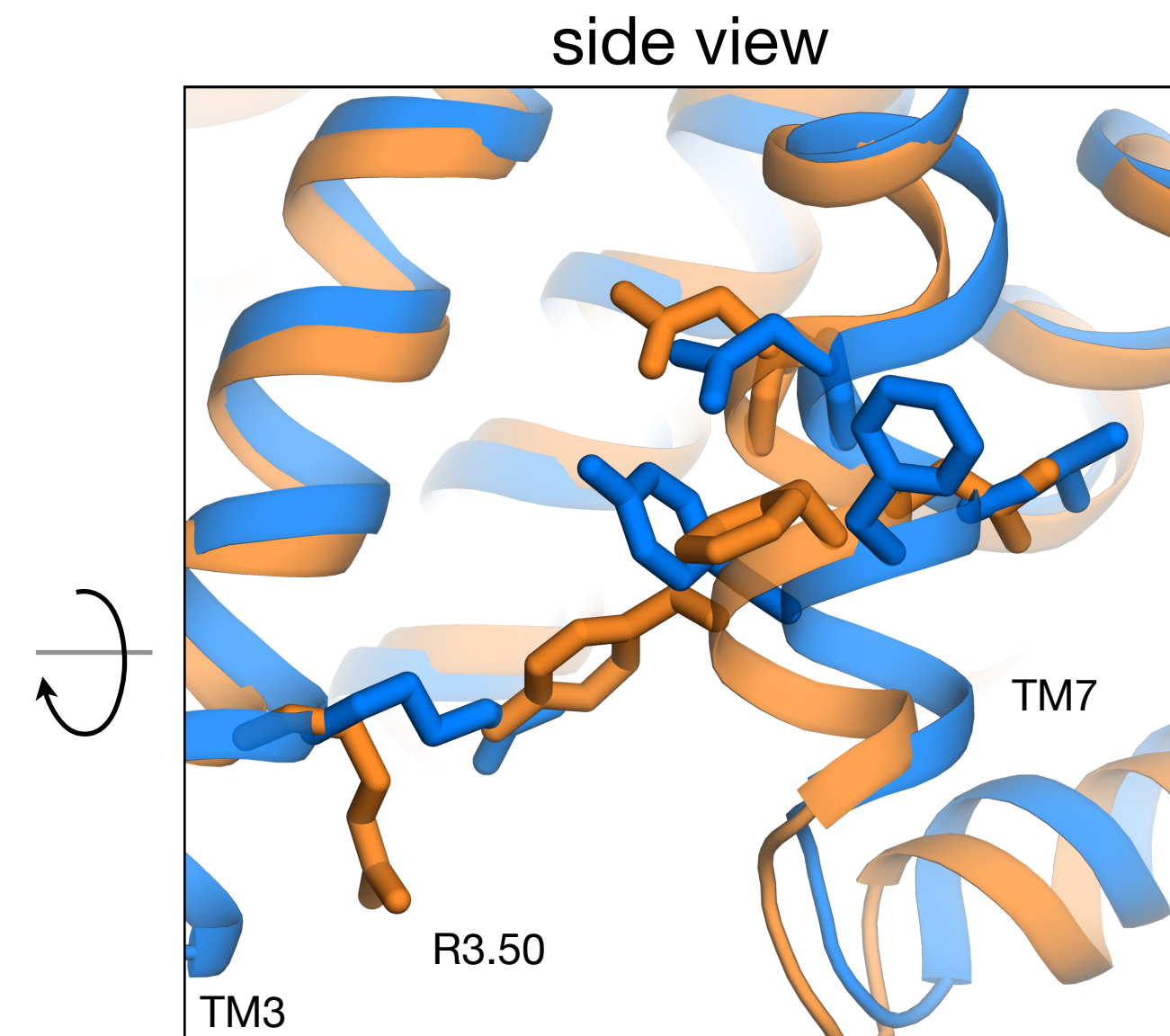
1. Simulate receptor **with no transducer bound**.
2. Identify conformations of the transducer coupling interface. Assess their potential to bind with G protein or arrestin.
3. Run simulations with agonists with a variety of bias profile and observe which receptor conformations they favor.
4. Confirm hypotheses by designing novel agonists or receptor mutations.



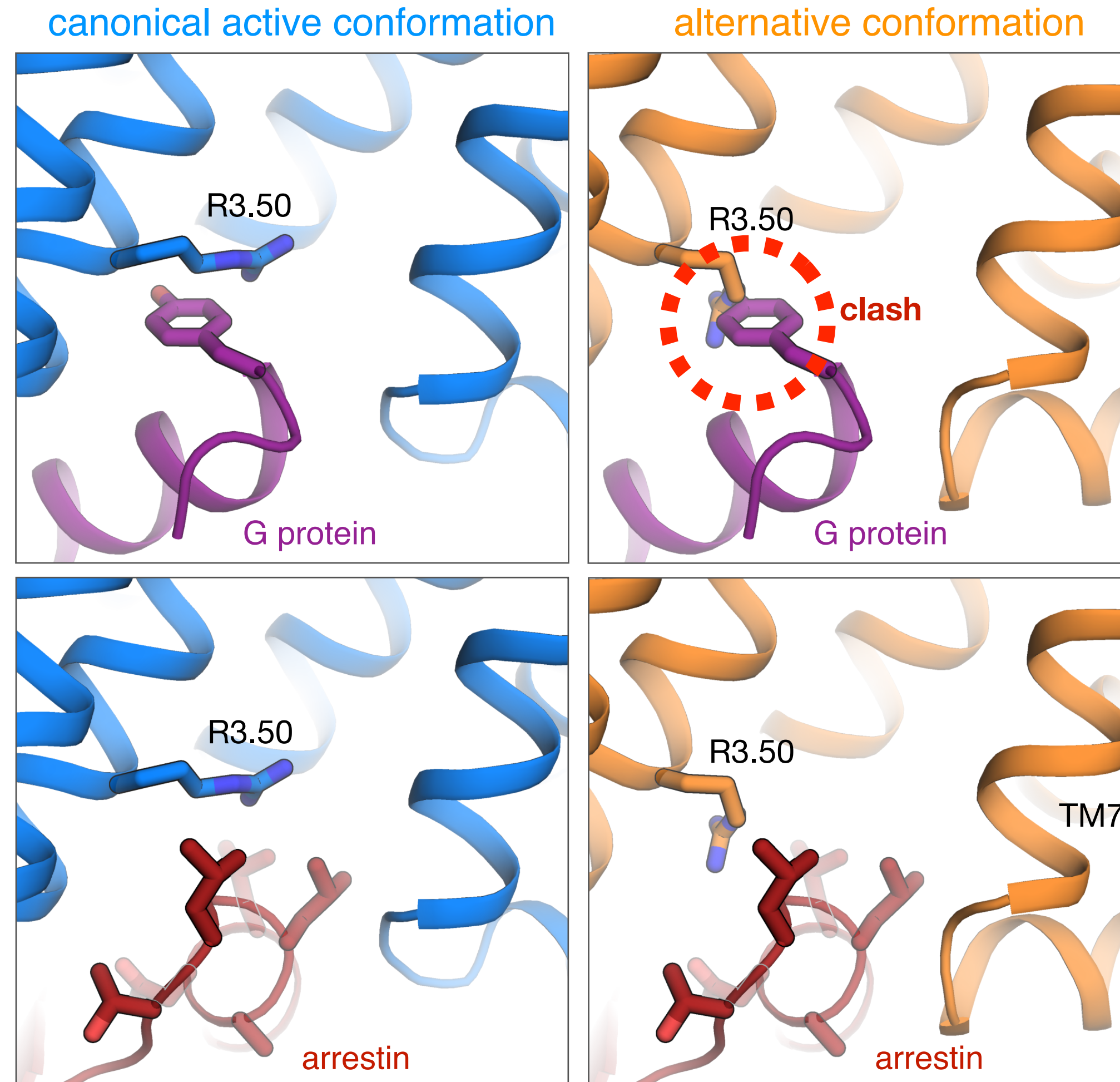
# Angiotensin receptor (AT<sub>1</sub>R) transitions between two active intracellular conformations in simulation



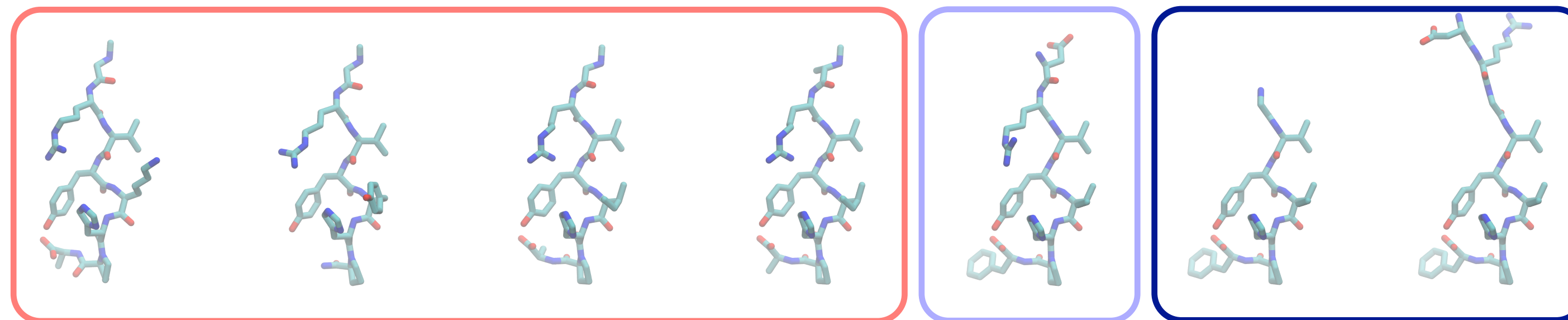
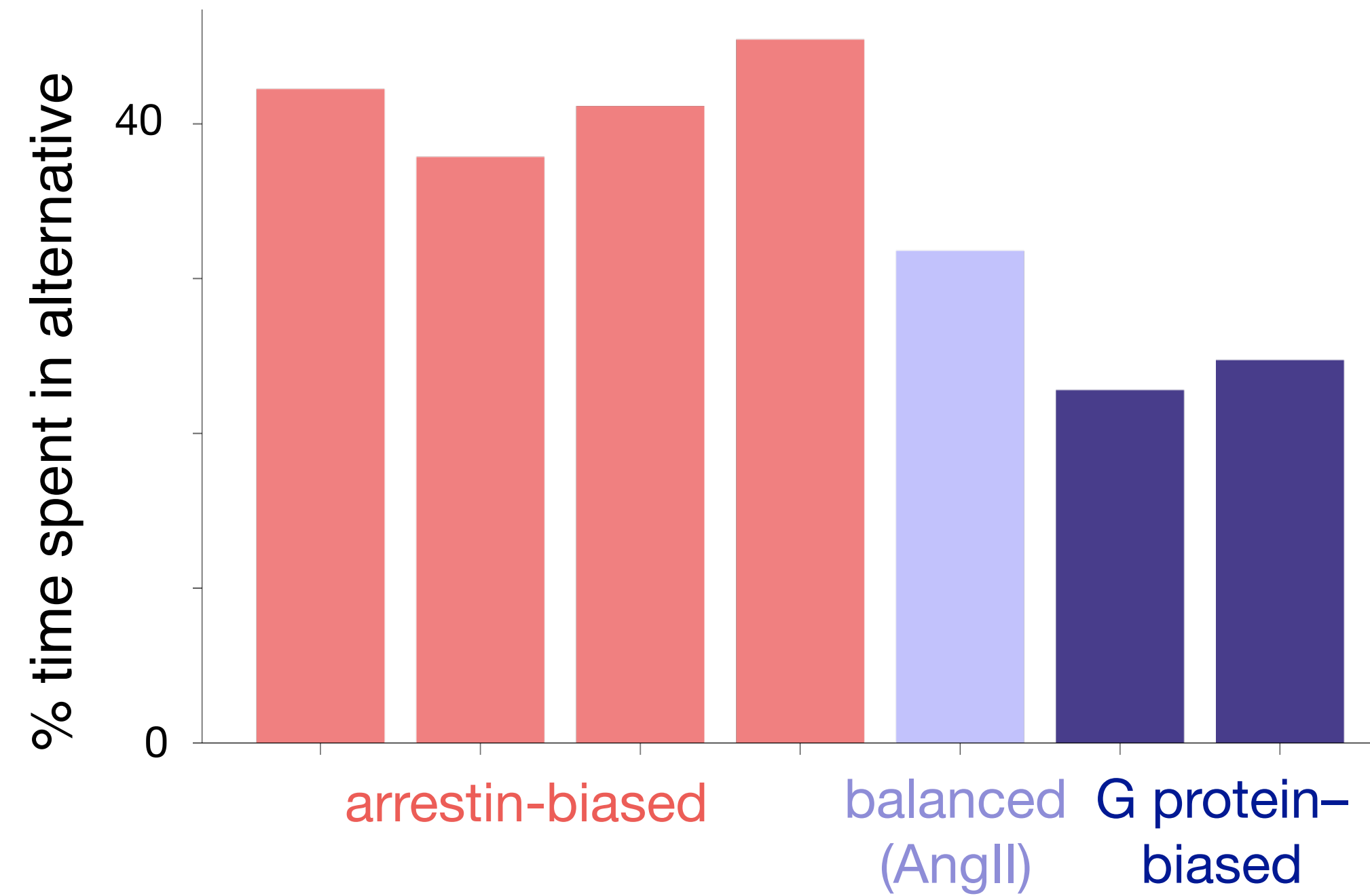
canonical active conformation  
alternative conformation



# Alternative conformation disfavors G-protein binding but can couple to arrestin



# Arrestin-biased ligands favor alternative conformation, G protein-biased ligands disfavor it



# Do these results transfer to opioid receptors?



Deniz Aydin

Article | [Published: 21 November 2022](#)

## Insights into distinct signaling profiles of the $\mu$ OR activated by diverse agonists

[Qianhui Qu](#), [Weijiao Huang](#), [Deniz Aydin](#), [Joseph M. Paggi](#), [Alpay B. Seven](#), [Haoqing Wang](#), [Soumen Chakraborty](#), [Tao Che](#), [Jeffrey F. DiBerto](#), [Michael J. Robertson](#), [Asuka Inoue](#), [Carl-Mikael Suomivuori](#), [Bryan L. Roth](#), [Susruta Majumdar](#) ✉, [Ron O. Dror](#) ✉, [Brian K. Kobilka](#) ✉ & [Georgios Skiniotis](#) ✉

[Nature Chemical Biology](#) (2022) | [Cite this article](#)



Yianni Laloudakis

Article | [Open Access](#) | [Published: 11 March 2023](#)

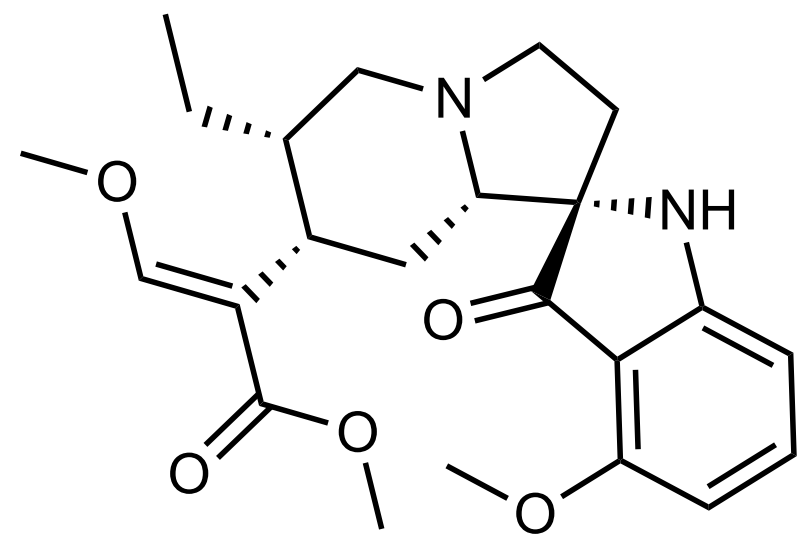
## Molecular mechanism of biased signaling at the kappa opioid receptor

[Amal El Daibani](#), [Joseph M. Paggi](#), [Kuglae Kim](#), [Yianni D. Laloudakis](#), [Petr Popov](#), [Sarah M. Bernhard](#), [Brian E. Krumm](#), [Reid H. J. Olsen](#), [Jeffrey Diberto](#), [F. Ivy Carroll](#), [Vsevolod Katritch](#), [Bernhard Wünsch](#), [Ron O. Dror](#) ✉ & [Tao Che](#) ✉

[Nature Communications](#) **14**, Article number: 1338 (2023) | [Cite this article](#)

# Agonists studied at $\mu$ OR

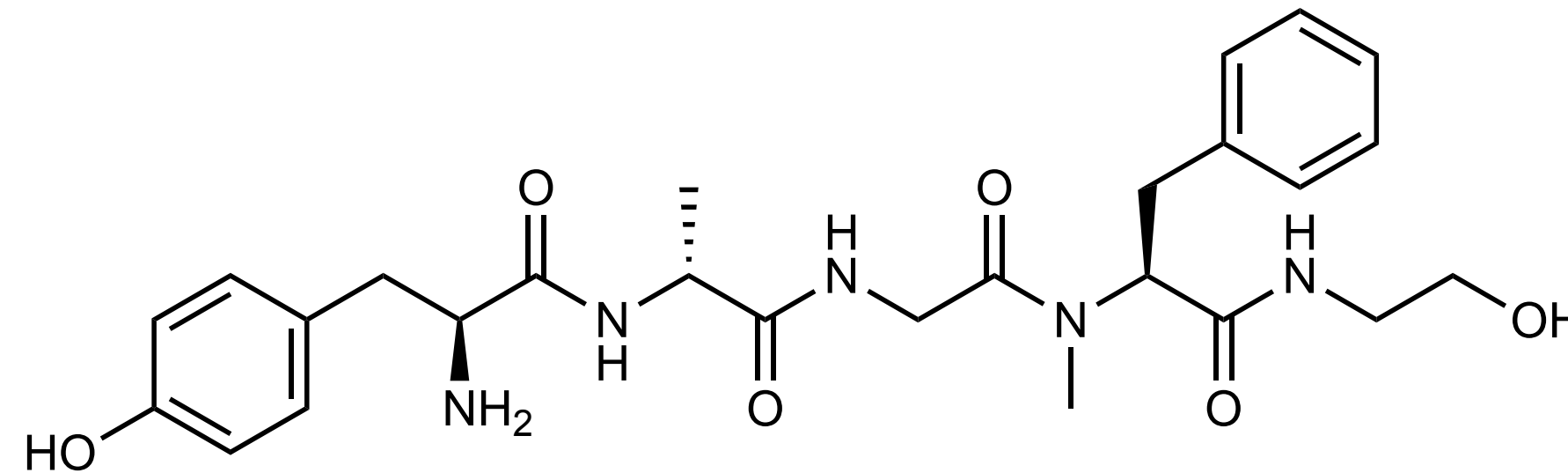
Mitragynine pseudoindoxyl (MP)



**G protein biased**

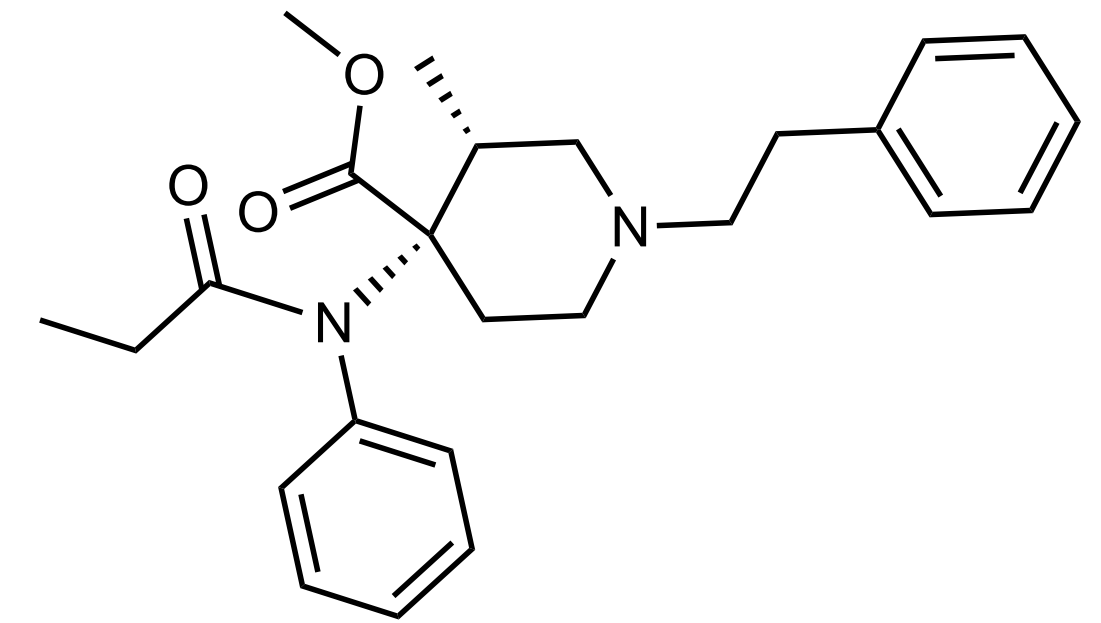


DAMGO



**Balanced**

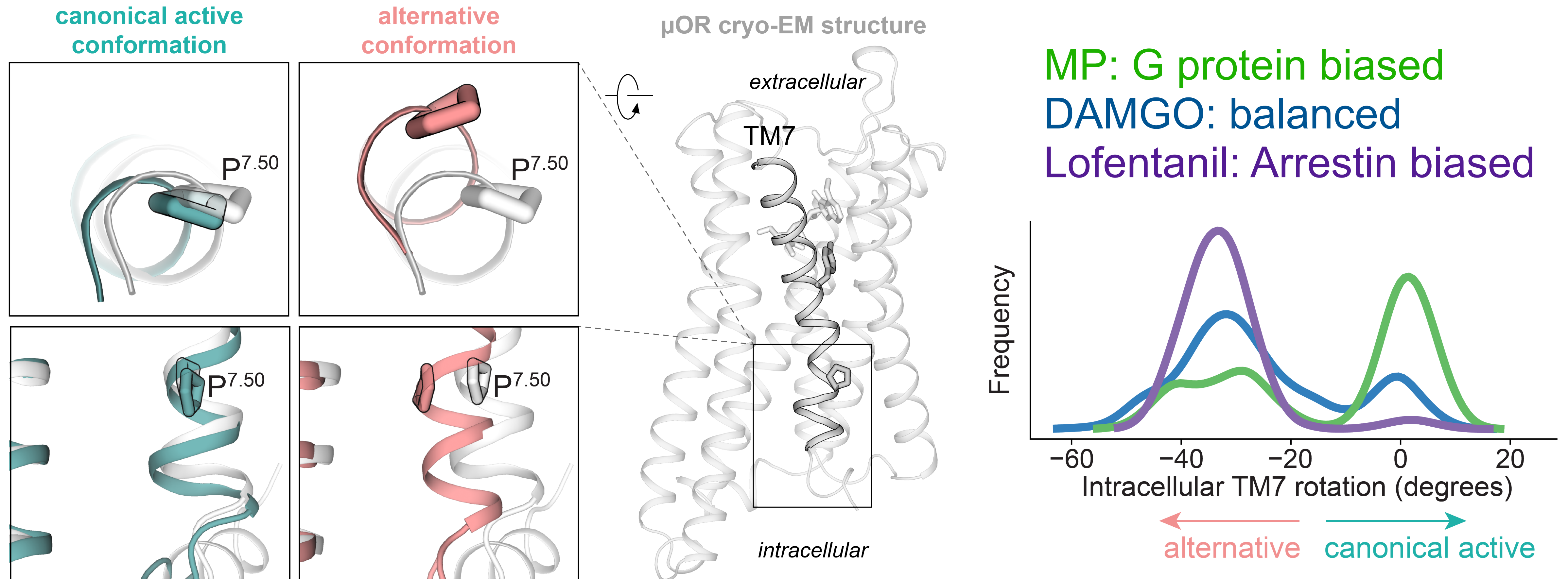
Lofentanil



**Arrestin biased**



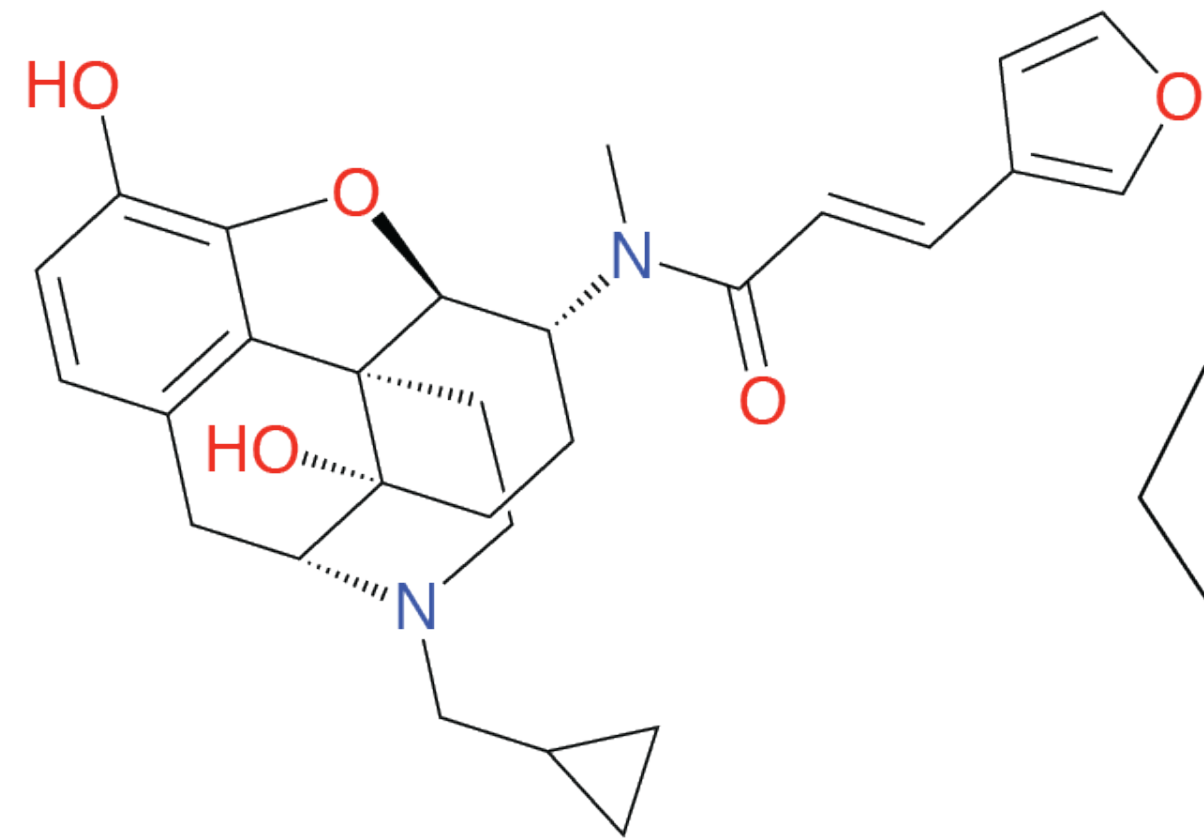
# We observe the canonical and alternative states at $\mu$ OR Occupancy of states explains bias profile of agonists



Statistical testing: Ran multiple independent simulations (6 for  $\mu$ OR, 10 for kOR), compute average value for each simulation, check for significance using t-test or Wilcoxon rank test.

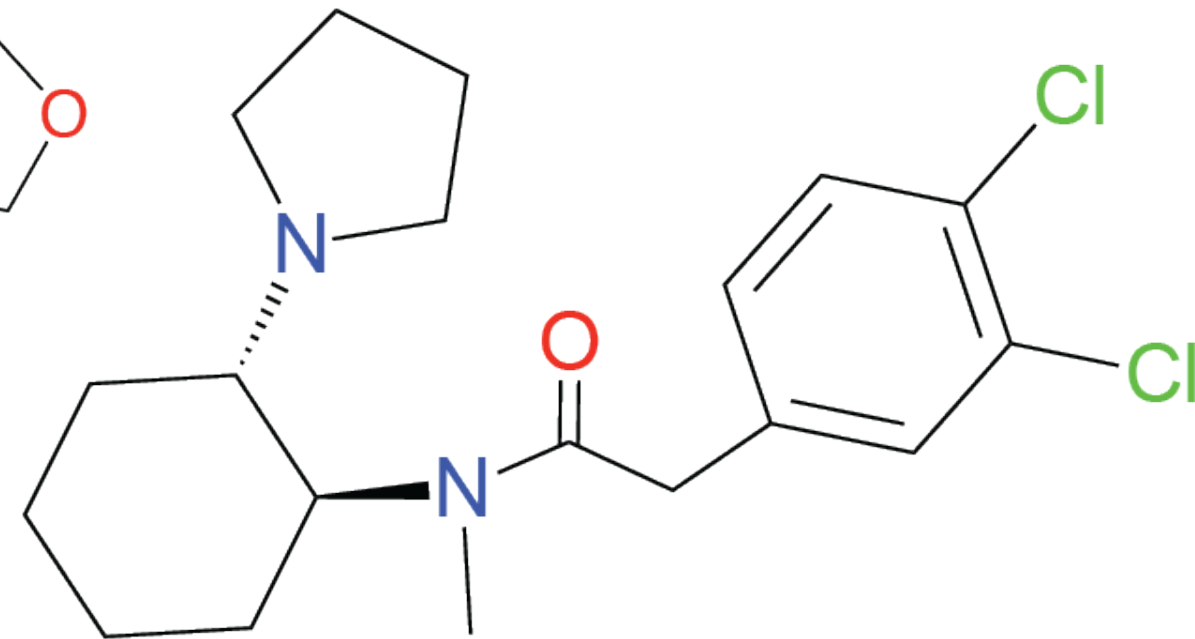
# Agonists studied at kOR

Nalfurafine



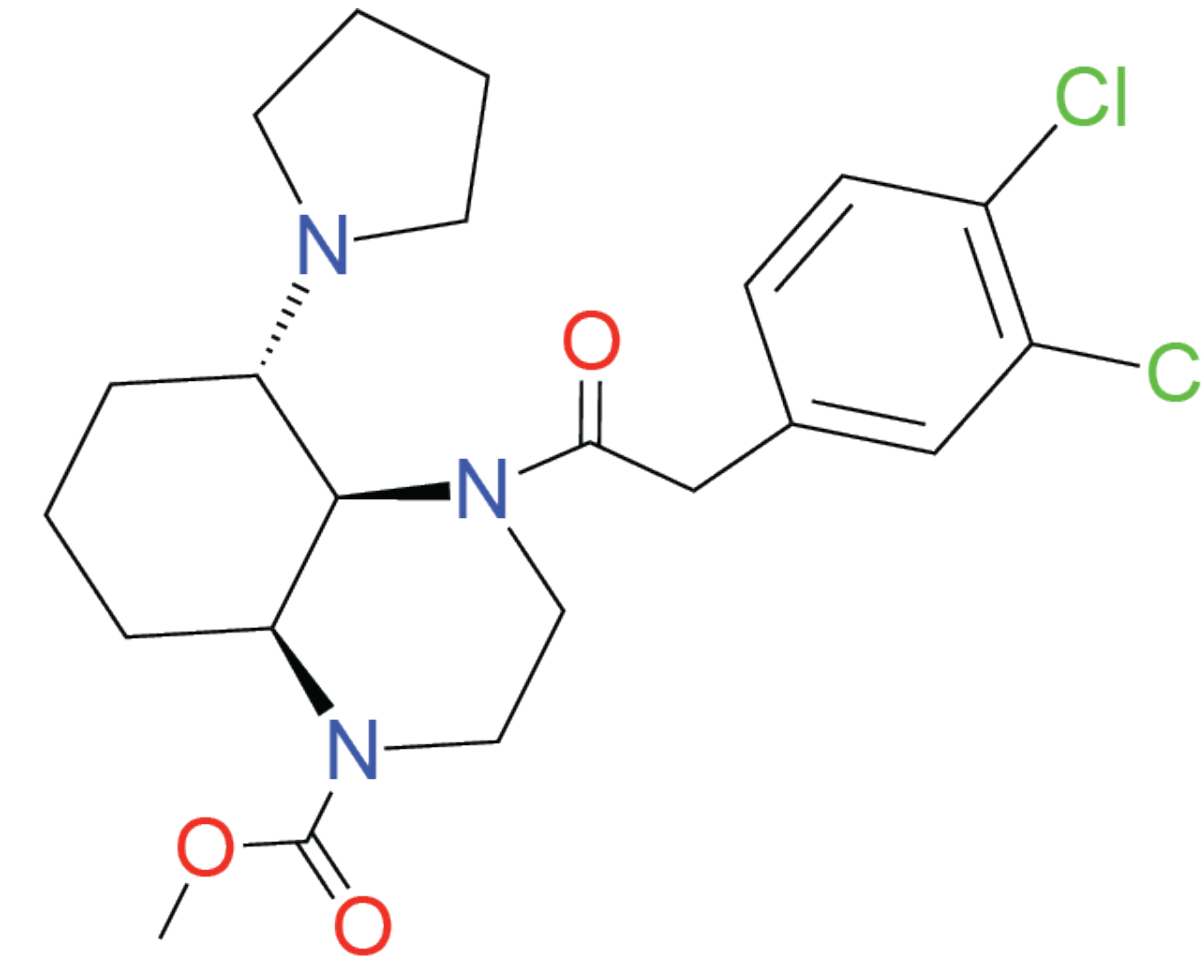
**G protein biased**

U50,488



**Balanced**

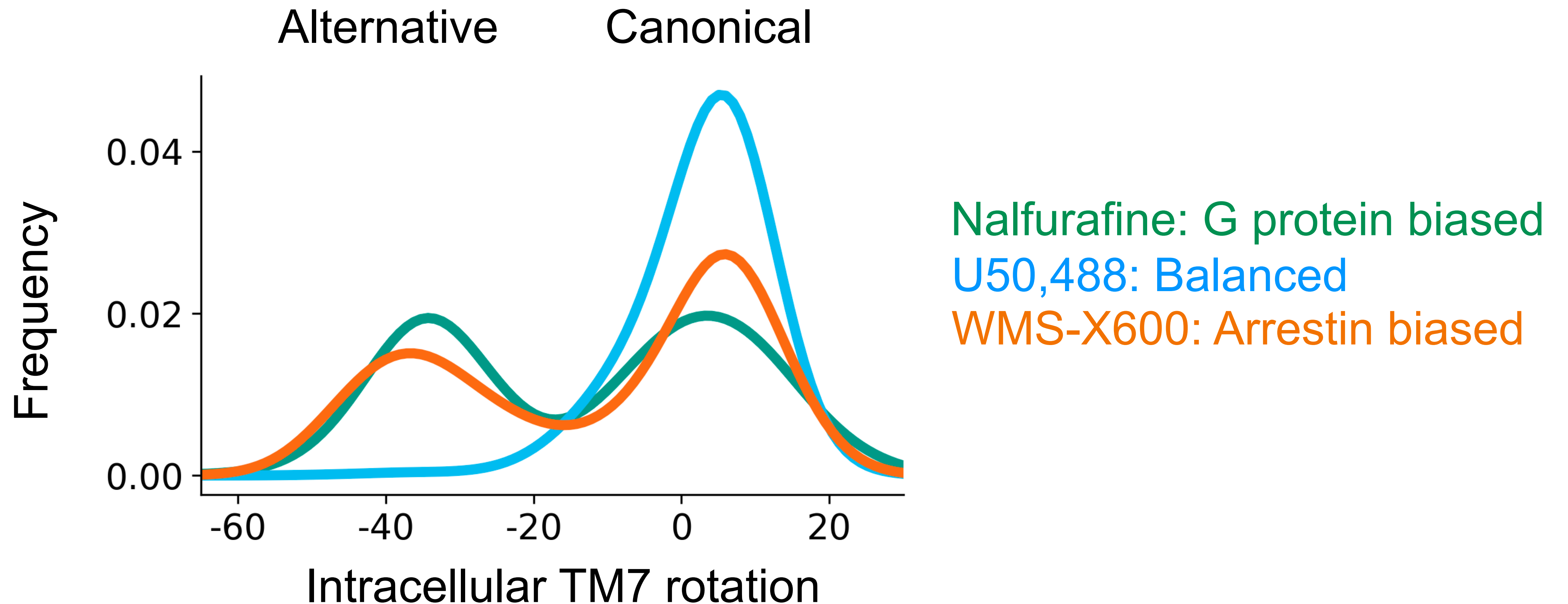
WMS-X600



**Arrestin biased**

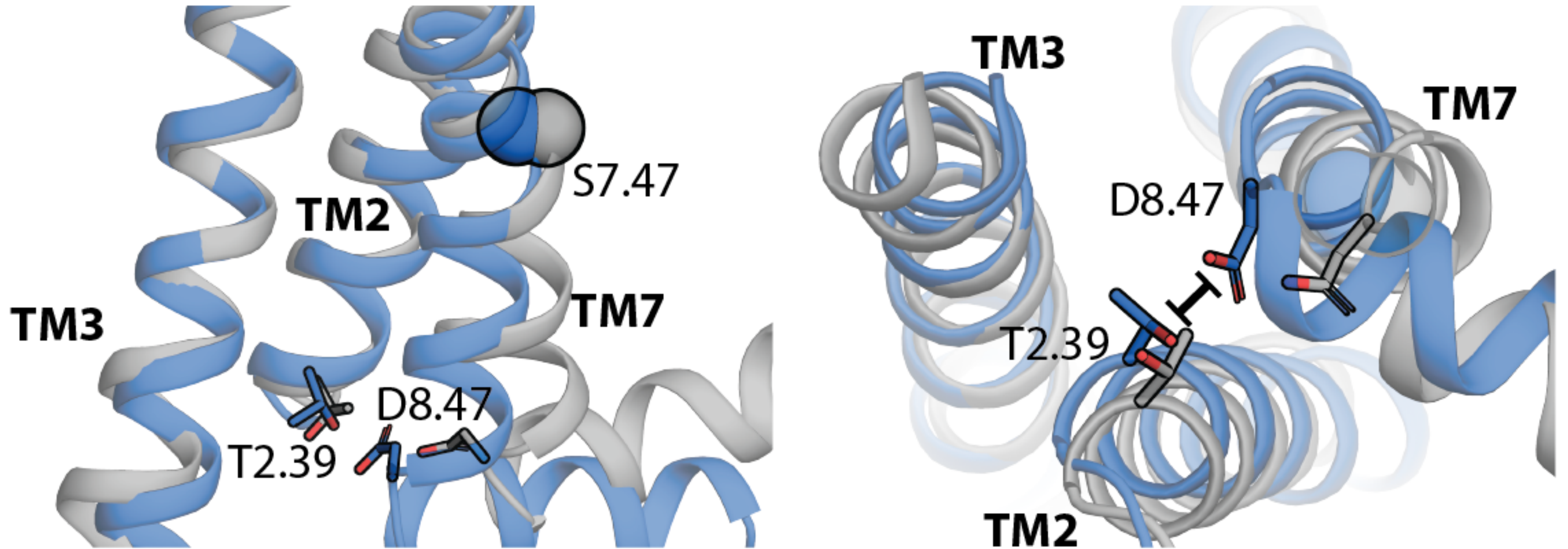
- Nalfurafine is approved in Japan since 2009 for use as an antipruritic, only one!
- Unlike other kOR agonists, Nalfurafine does not induce dysphoria at therapeutic doses
- Believed to be at least in part due to G protein bias

# The balanced agonist already maxes out canonical state. How can you get G protein bias?

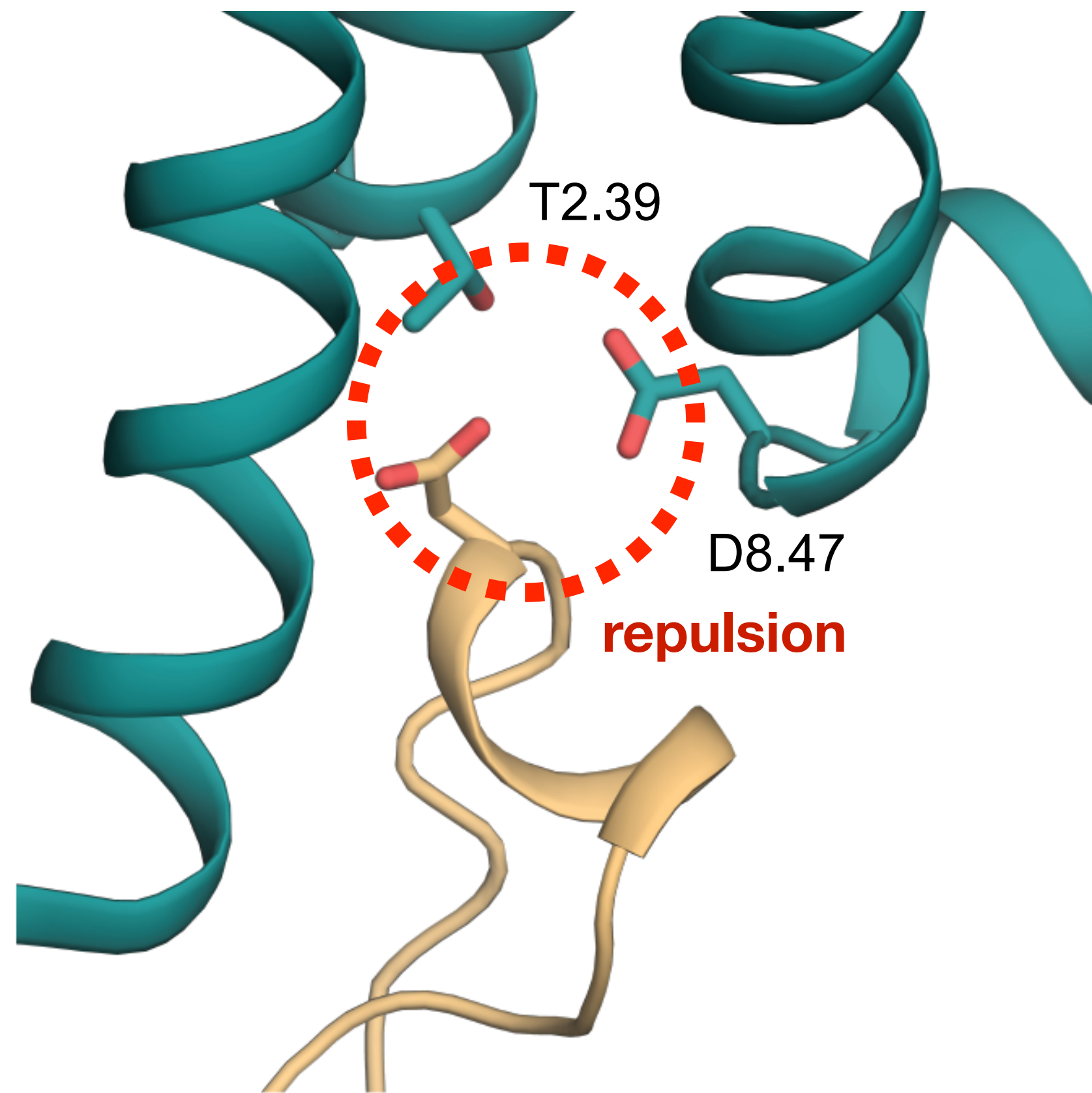




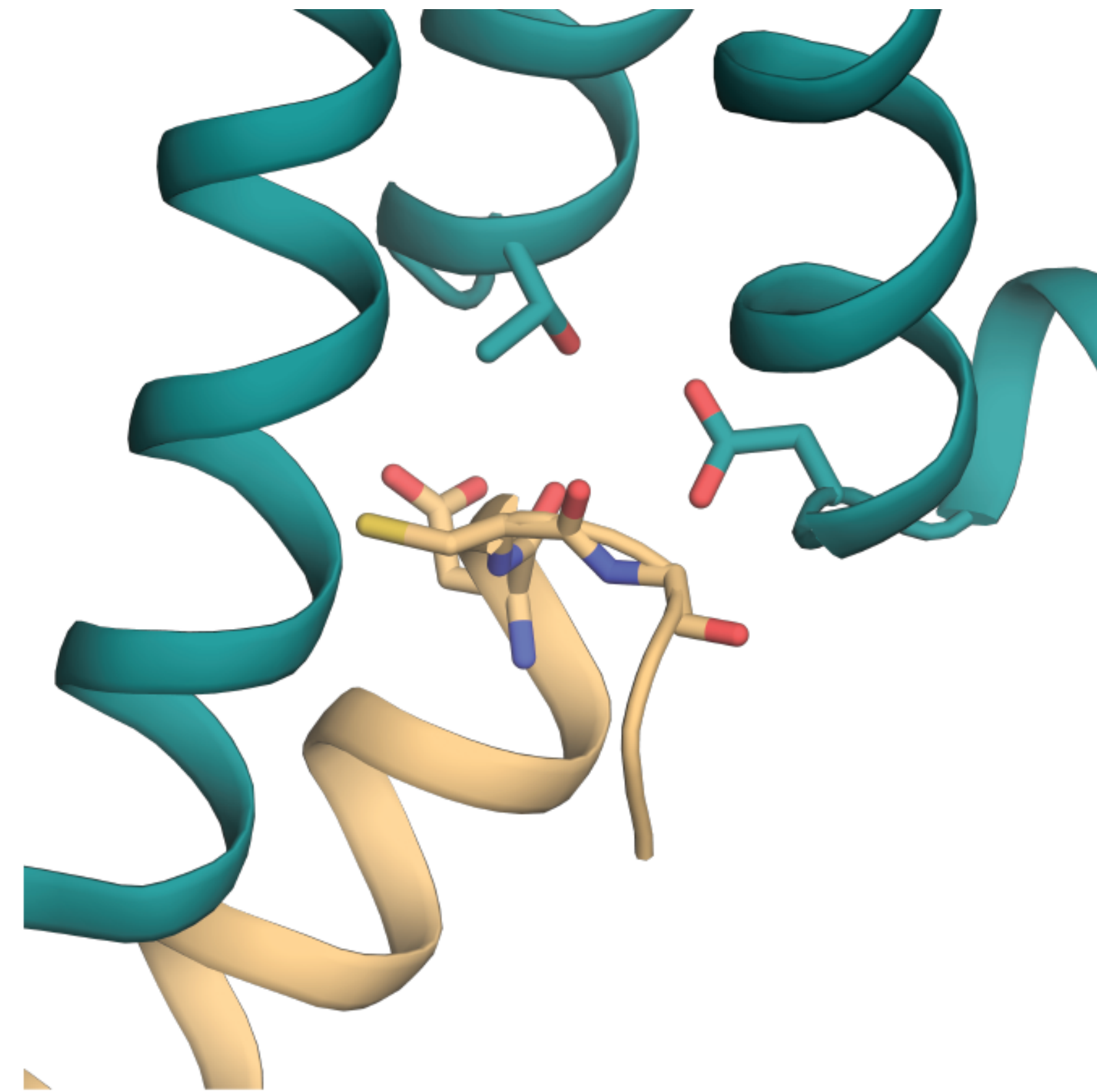
At kOR, we observe a third receptor conformation: the “occluded state”



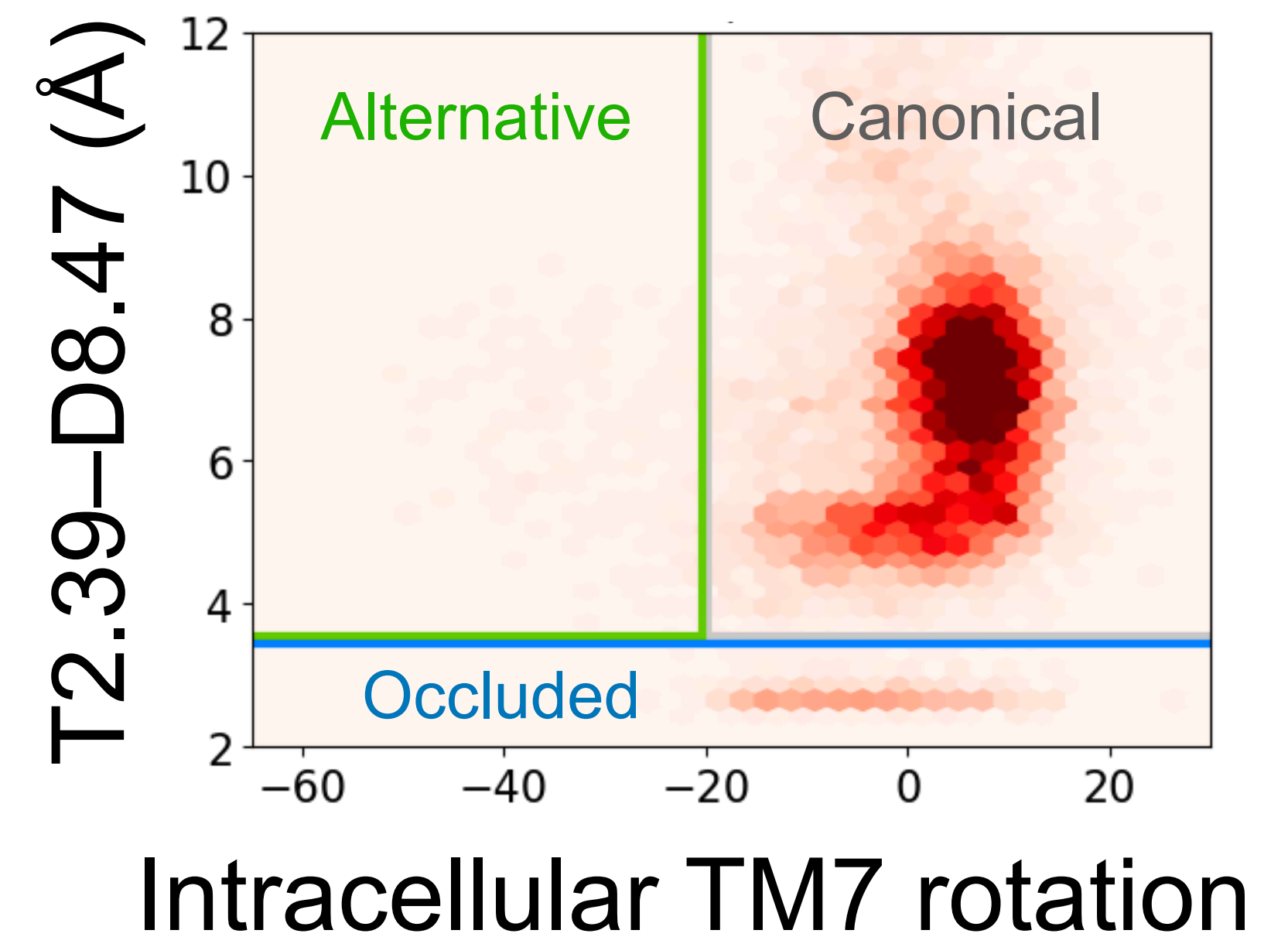
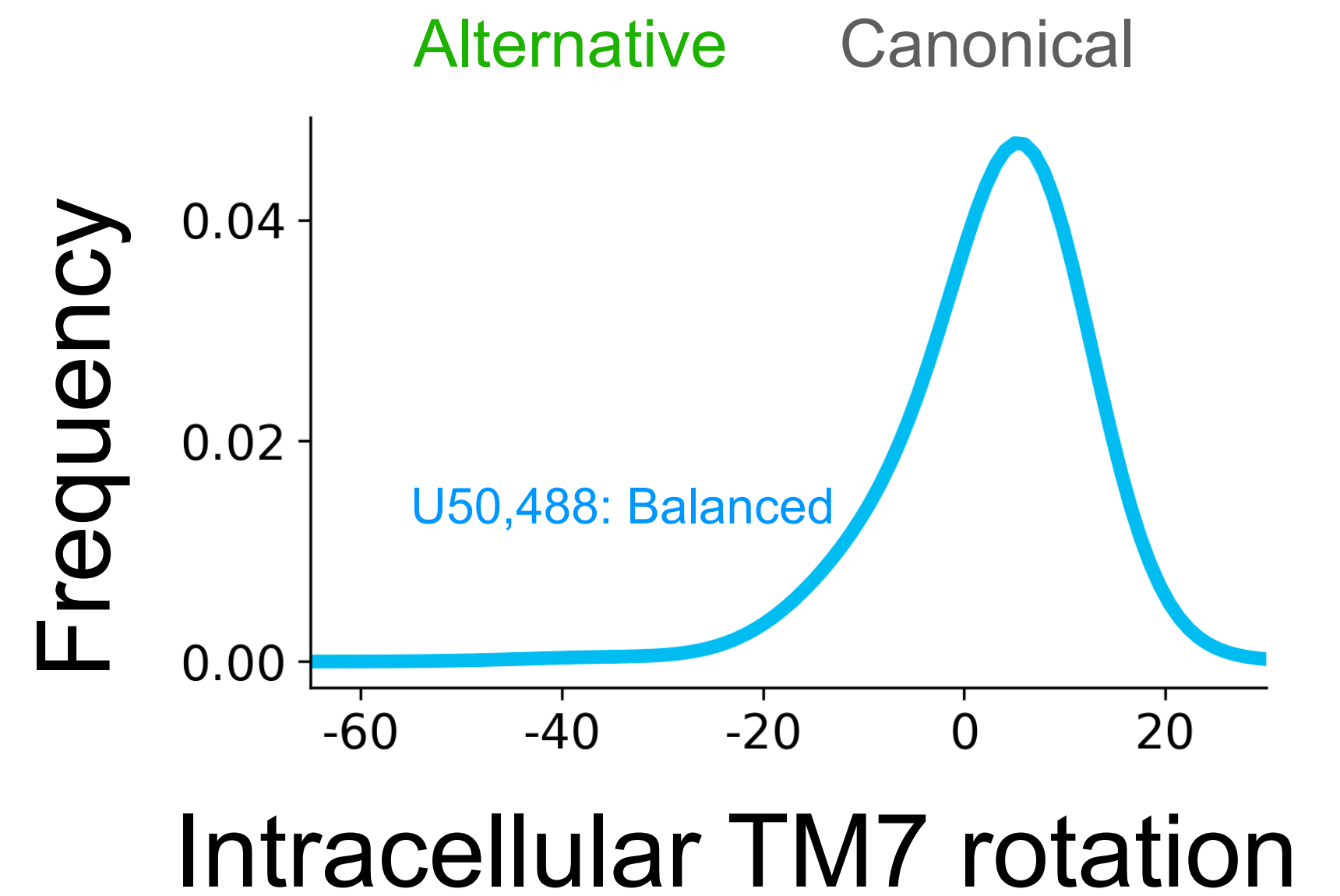
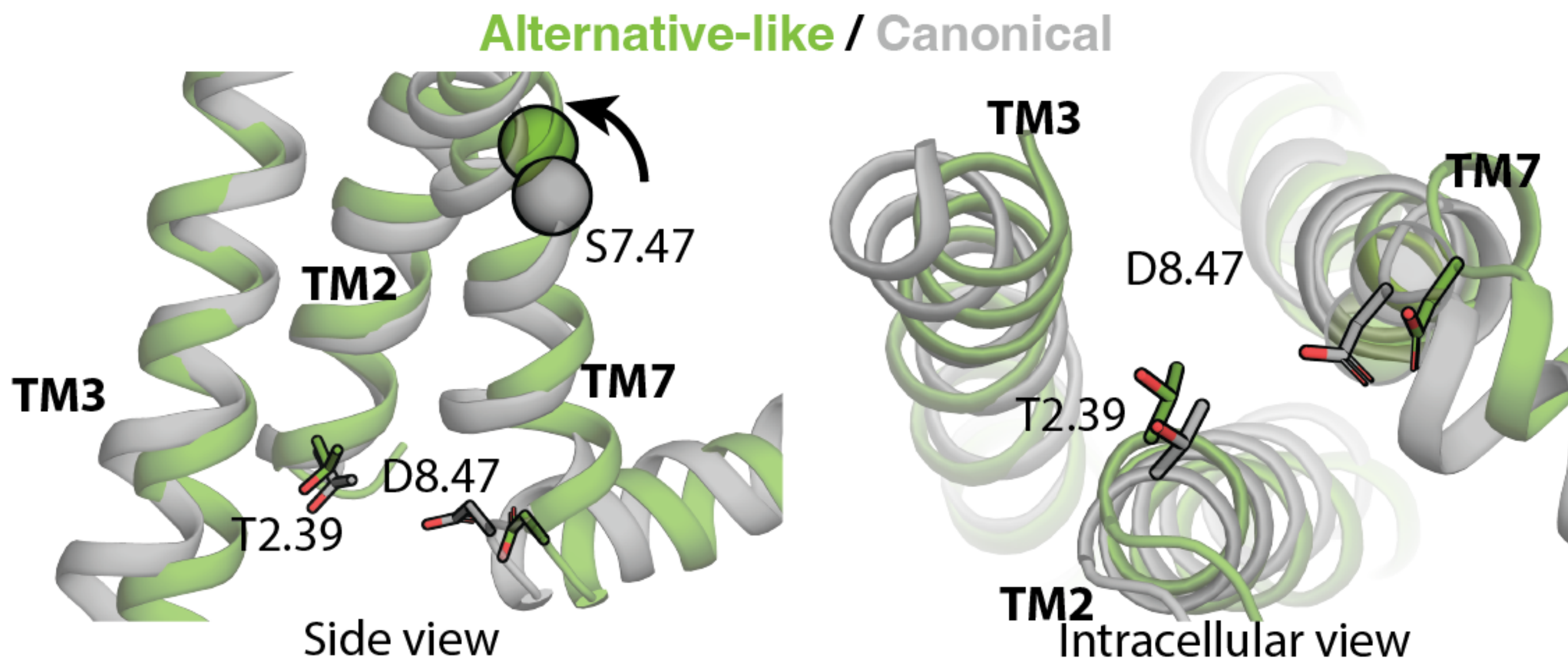
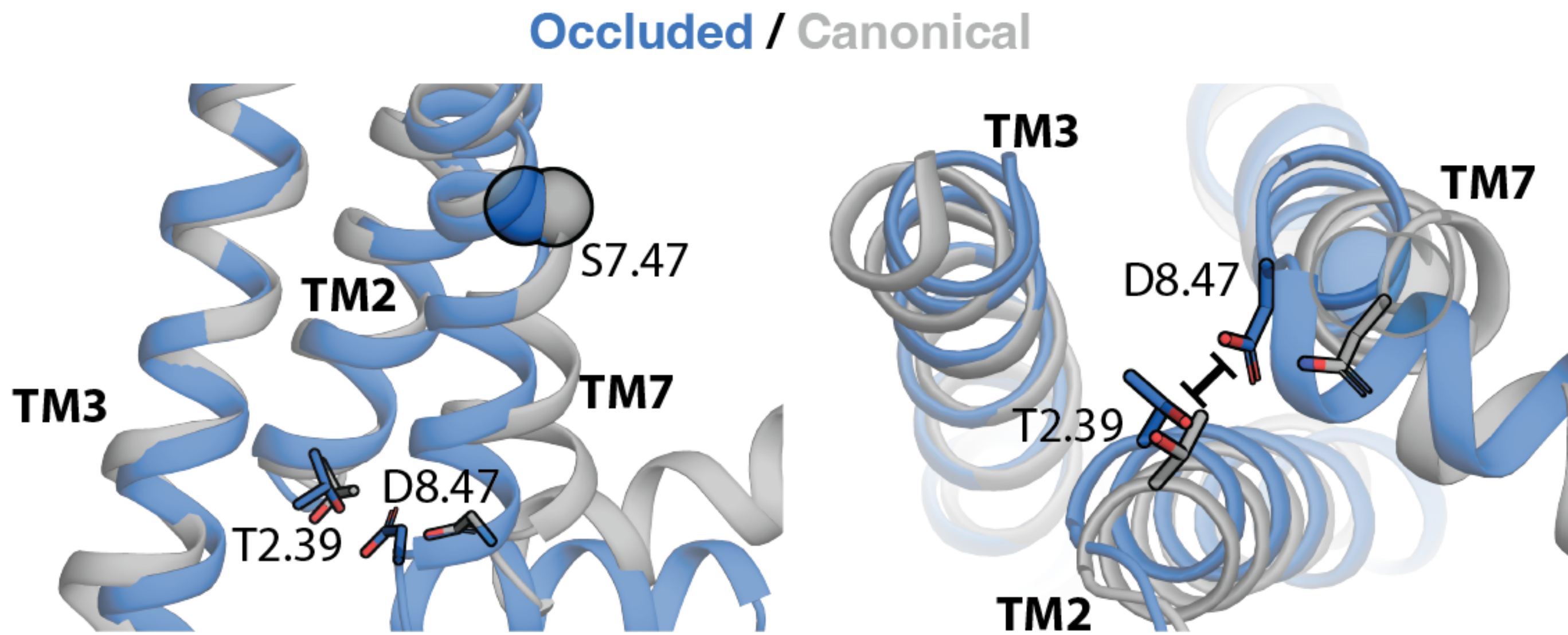
# The occluded state presents an electrostatic barrier to arrestin coupling



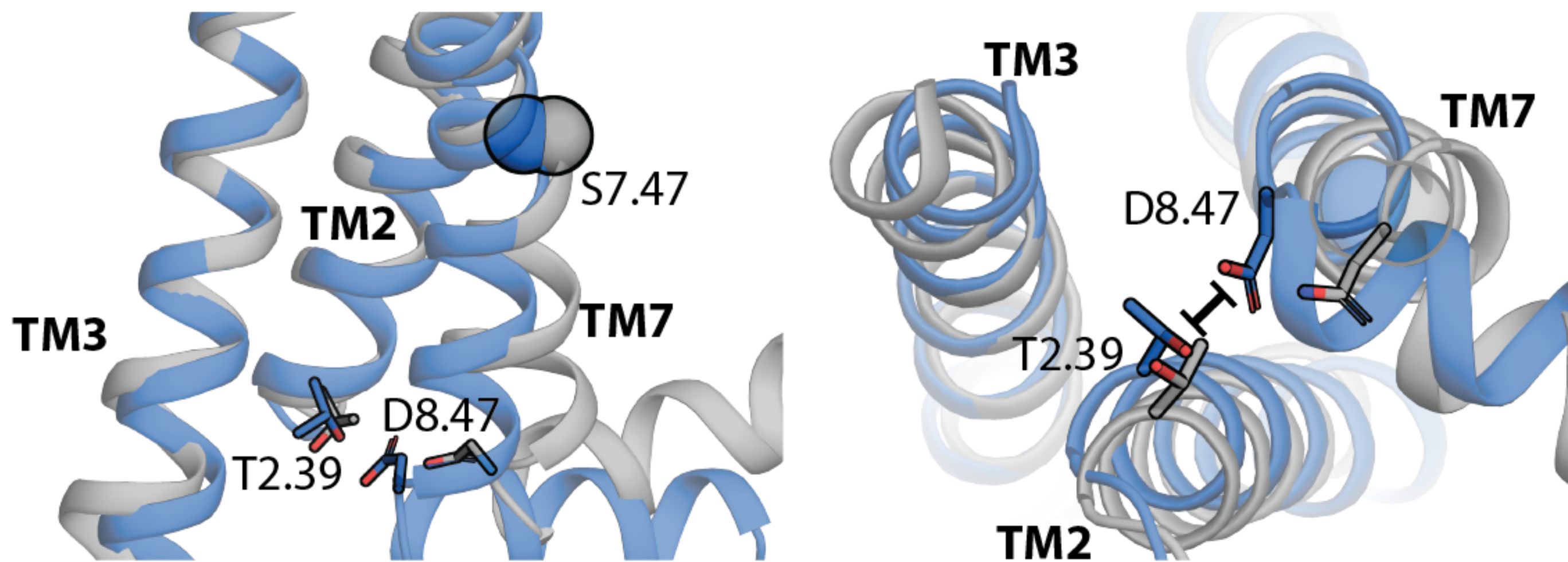
kOR occluded state  
NTSR1–arrestin-2



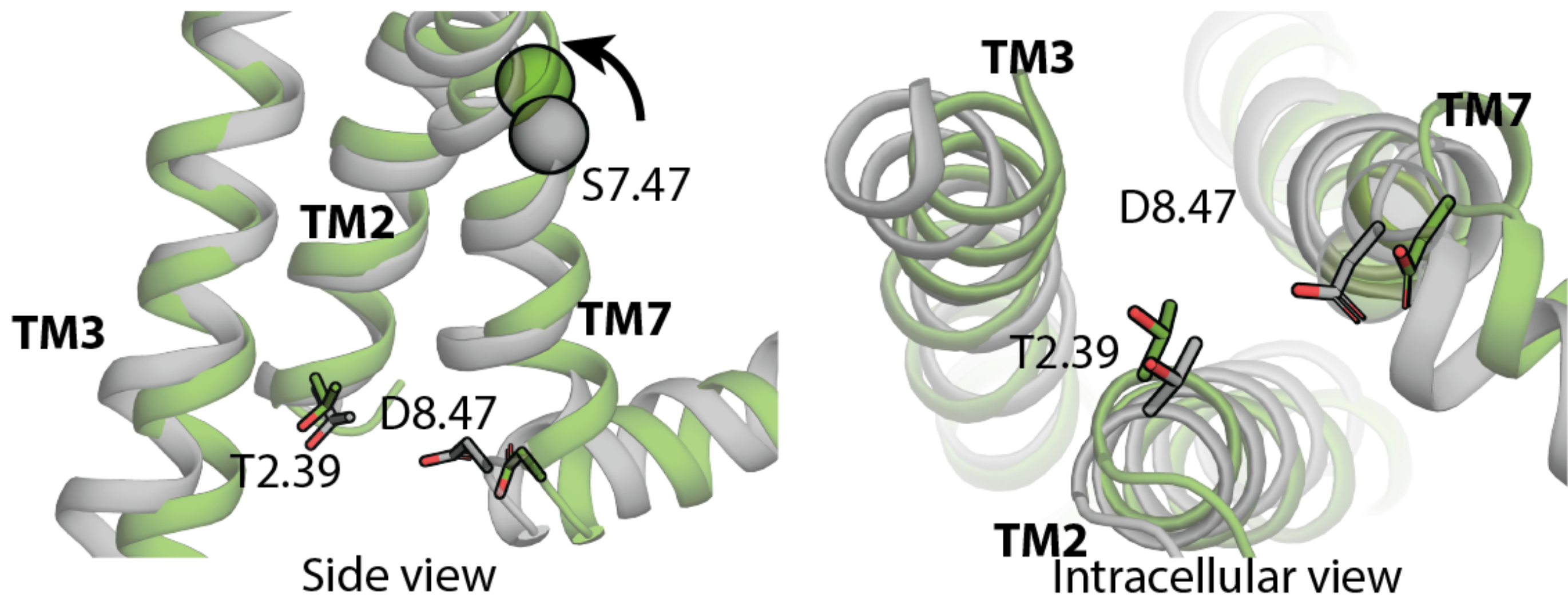
kOR occluded state  
 $\mu$ OR–Gi



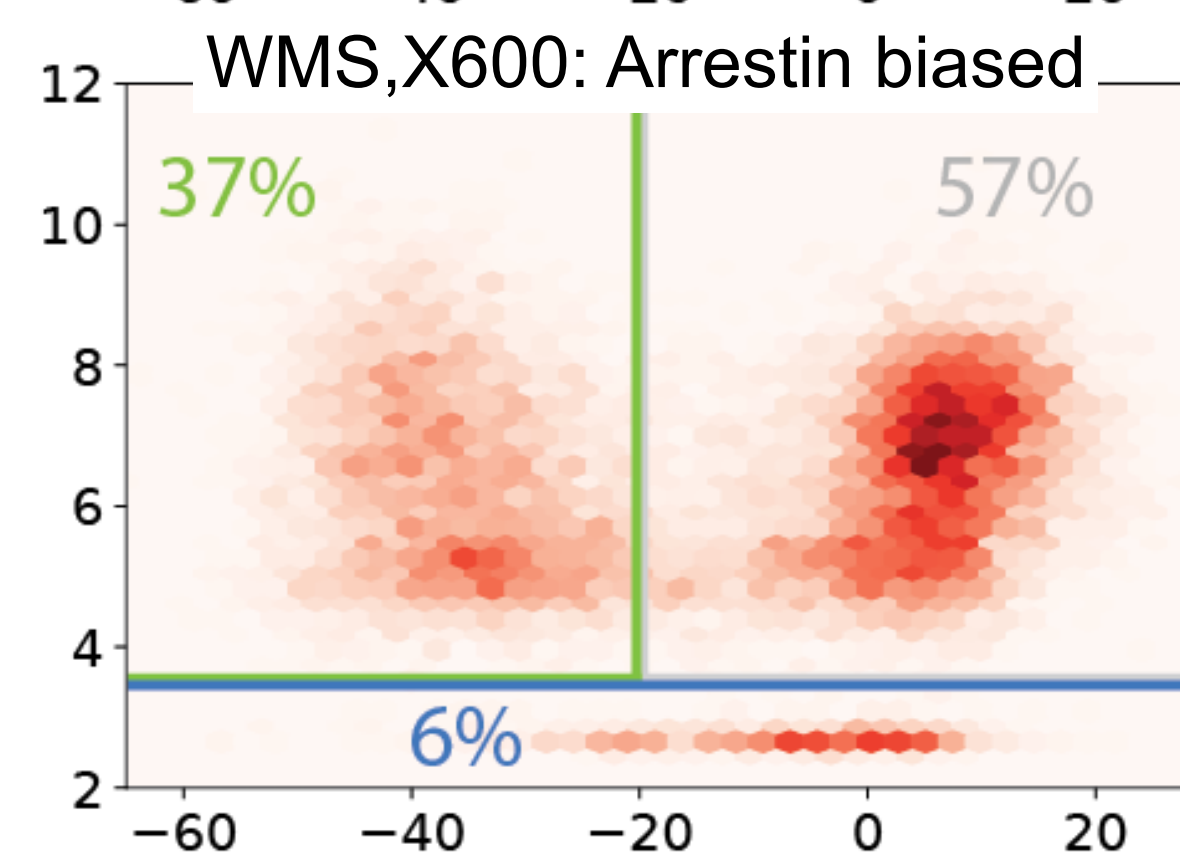
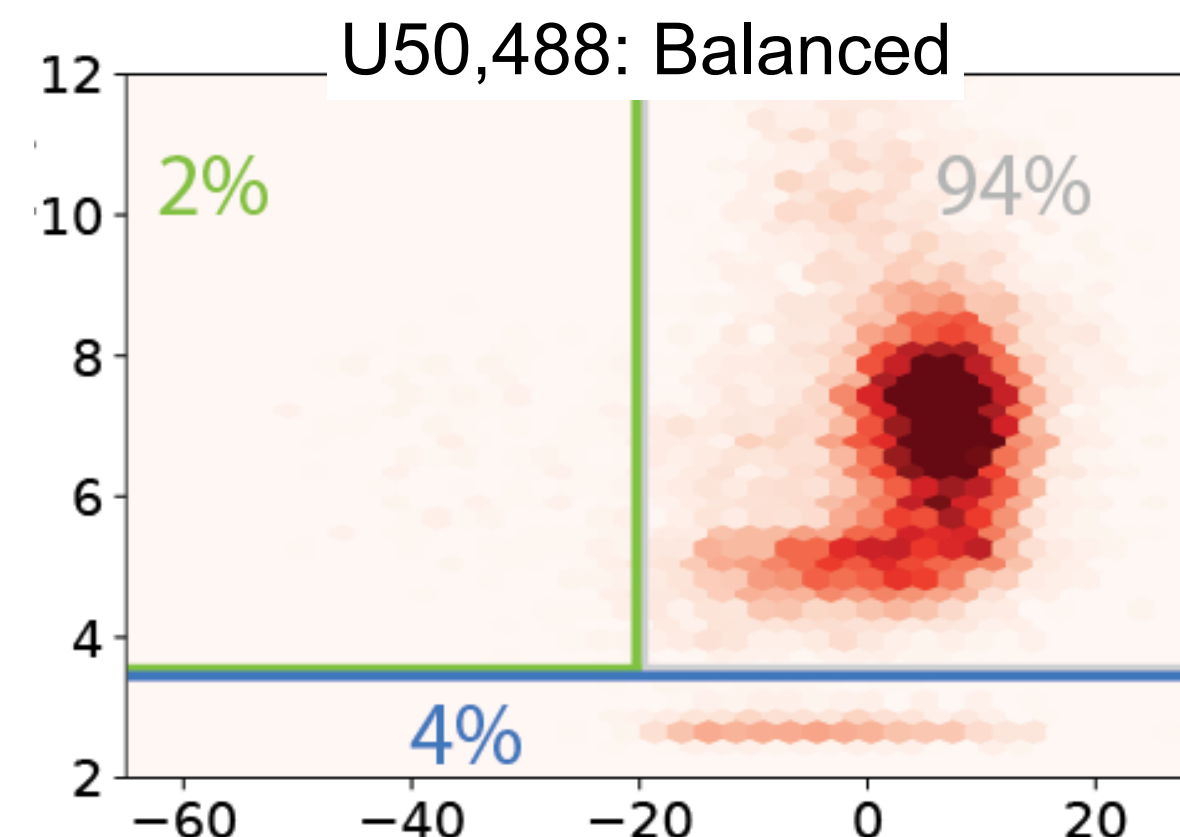
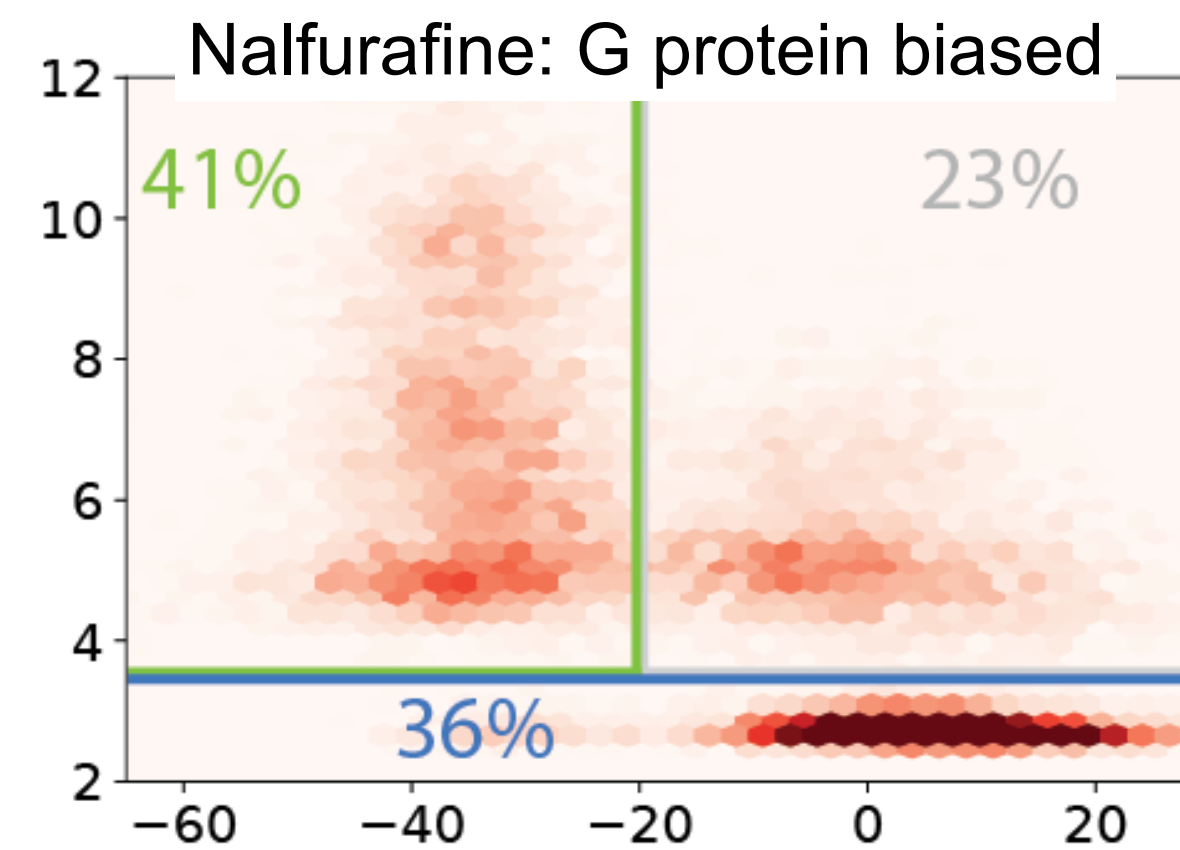
Occluded / Canonical



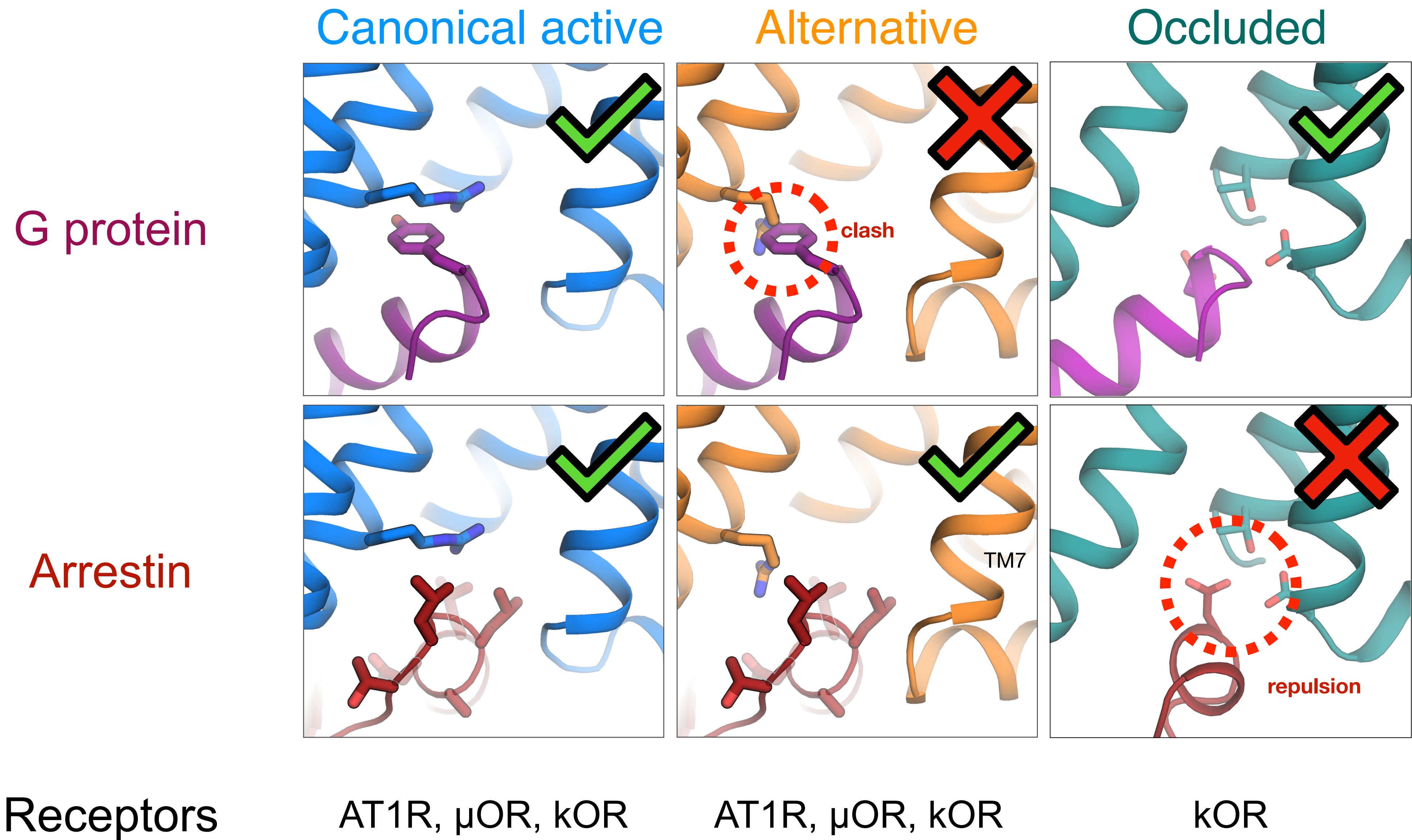
Alternative-like / Canonical



T2.39-D8.47 (Å)



Intracellular TM7 rotation



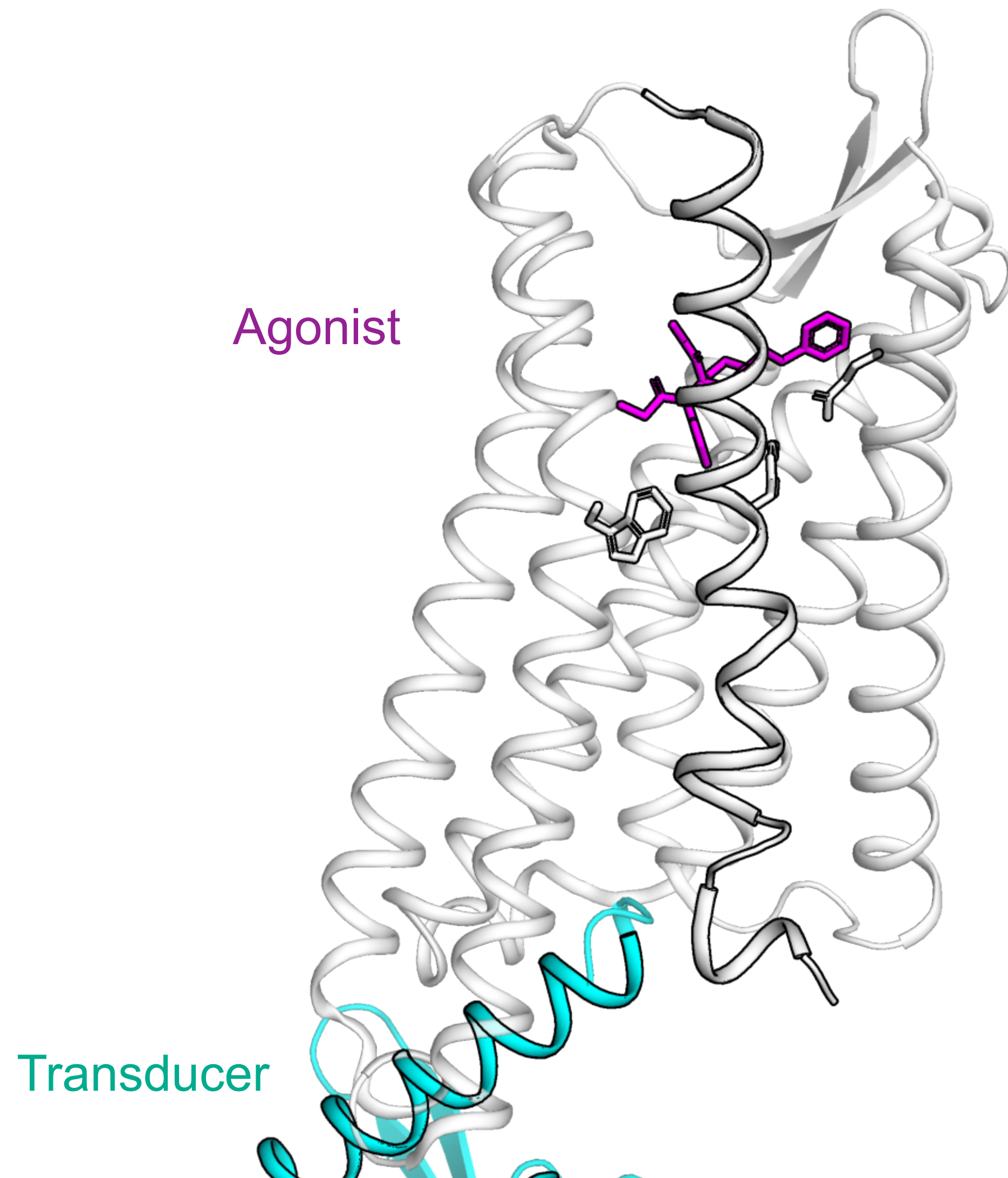
... ?

1. What are these “biased” conformations?

**2. How do agonists favor different conformations?**

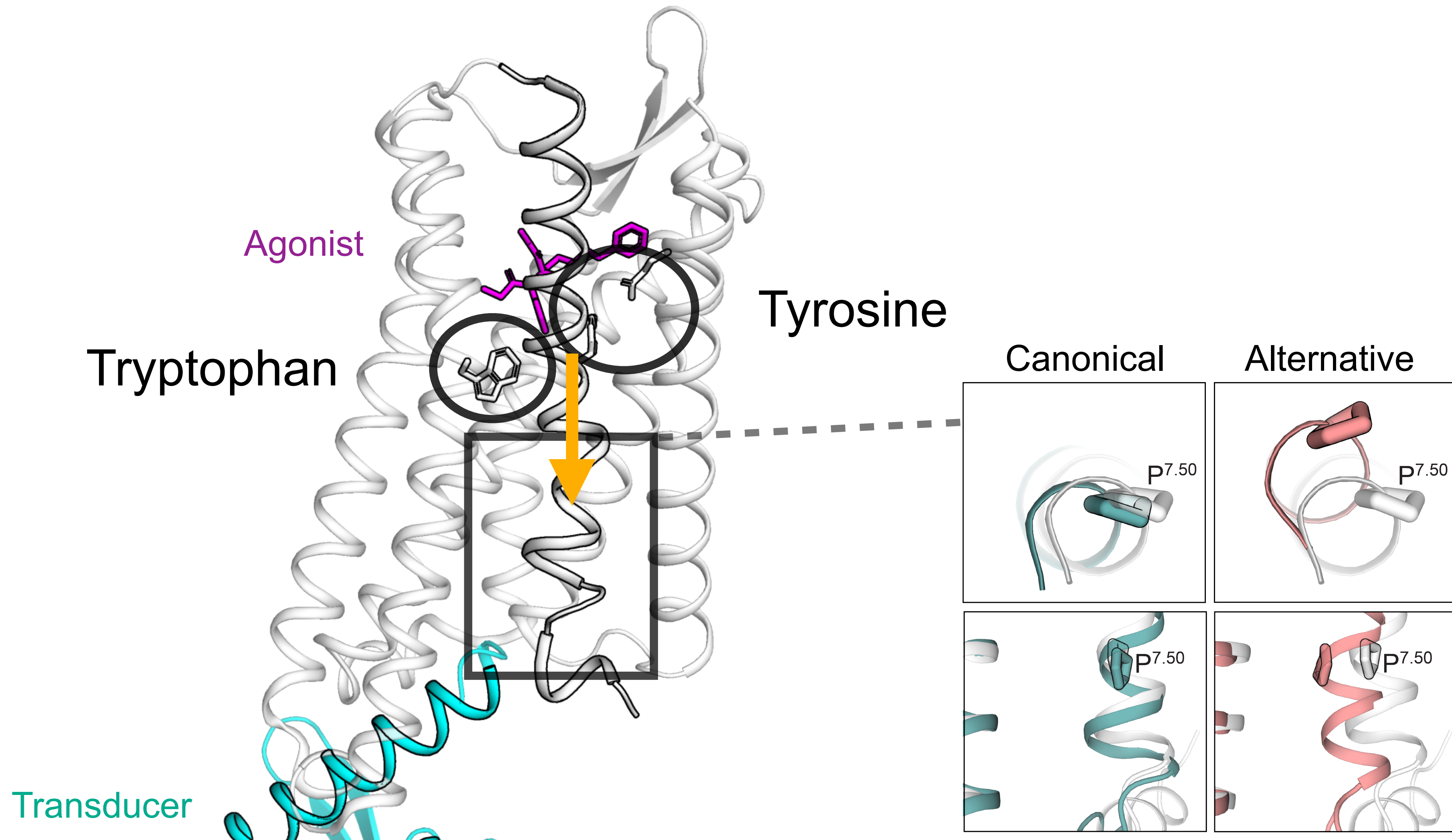
- What are the differences in protein–ligand interactions?
- How are these differences transmitted through the receptor?

# Before we get lost in the trees...



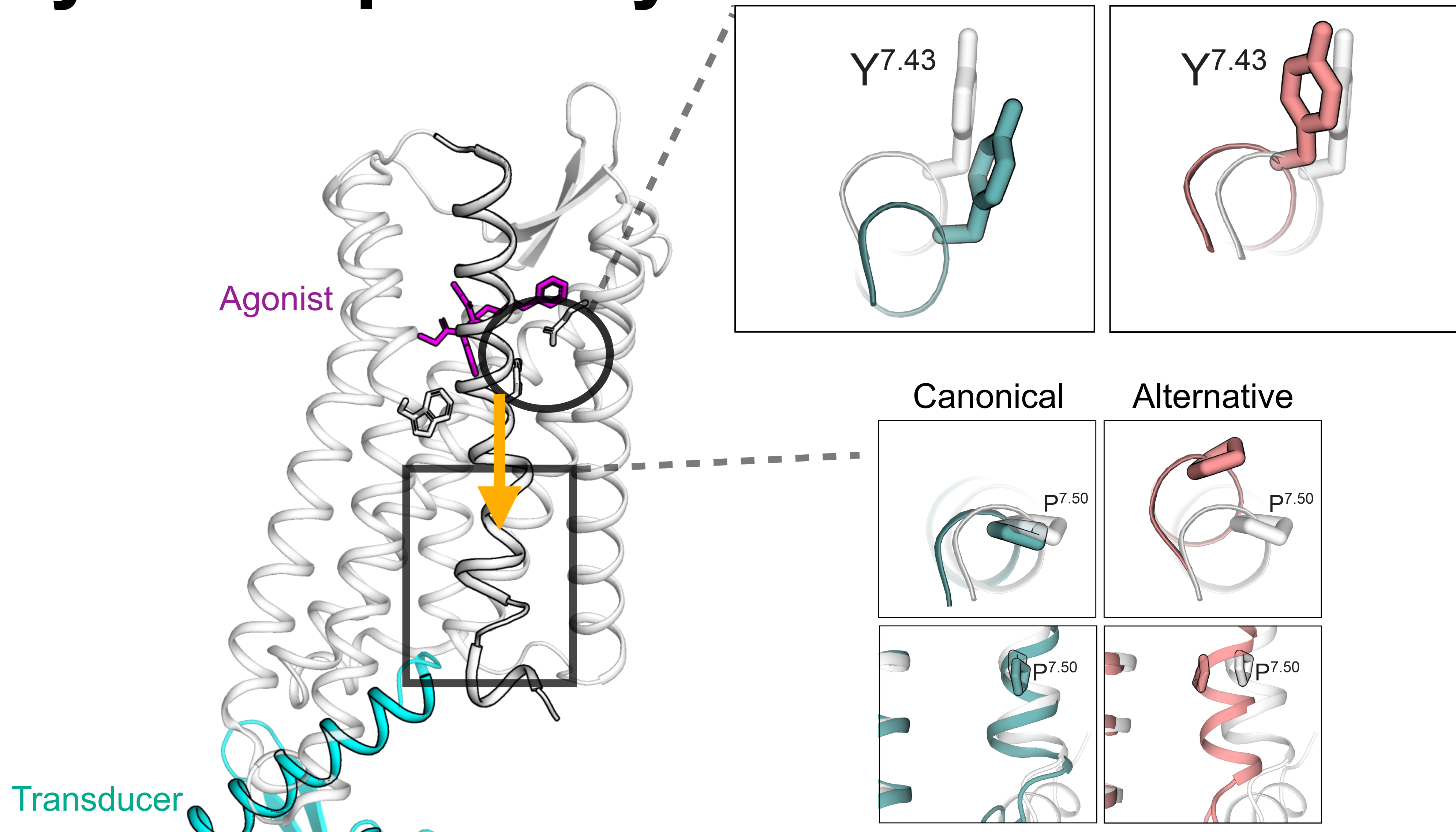
- I'm only going to share a subset of the results here
  - See our papers for more details!
  - We validate much of our proposed mechanism with mutagenesis experiments
- 1. Multiple layers of abstraction, things get confusing if you stay too low
  - Transducer site conformations
  - Allosteric pathways
  - Direct protein–ligand interactions
- 2. MD is a powerful tool
  - Subtle differences in ligands can have large and hard to predict impacts on binding pocket conformations
  - Even for major differences between ligands, the implications on protein dynamics are not clear from structures alone

**Most importantly, a rigid body rotation connects the binding pocket to the transducer binding site**



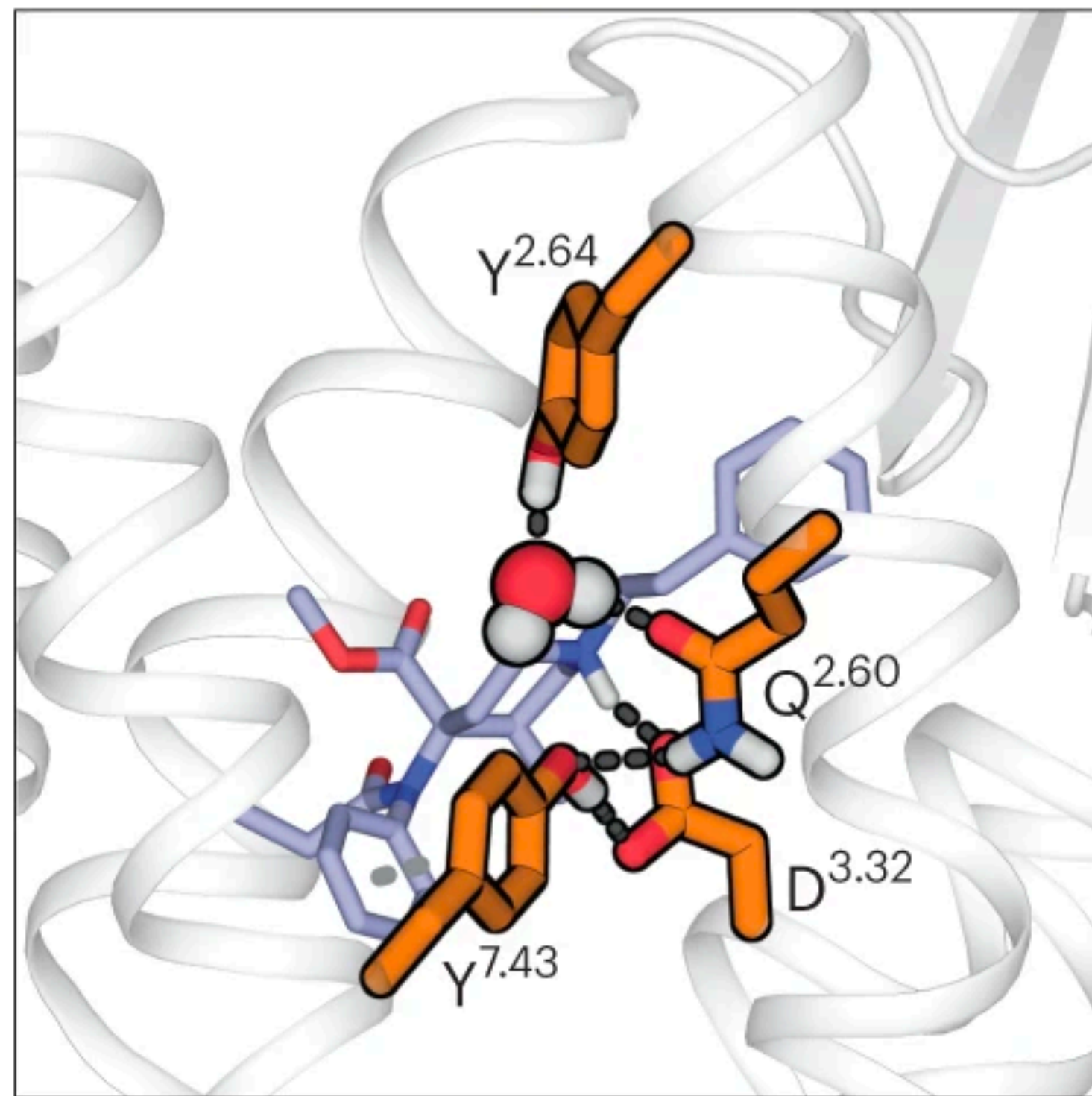


# Tyrosine pathway

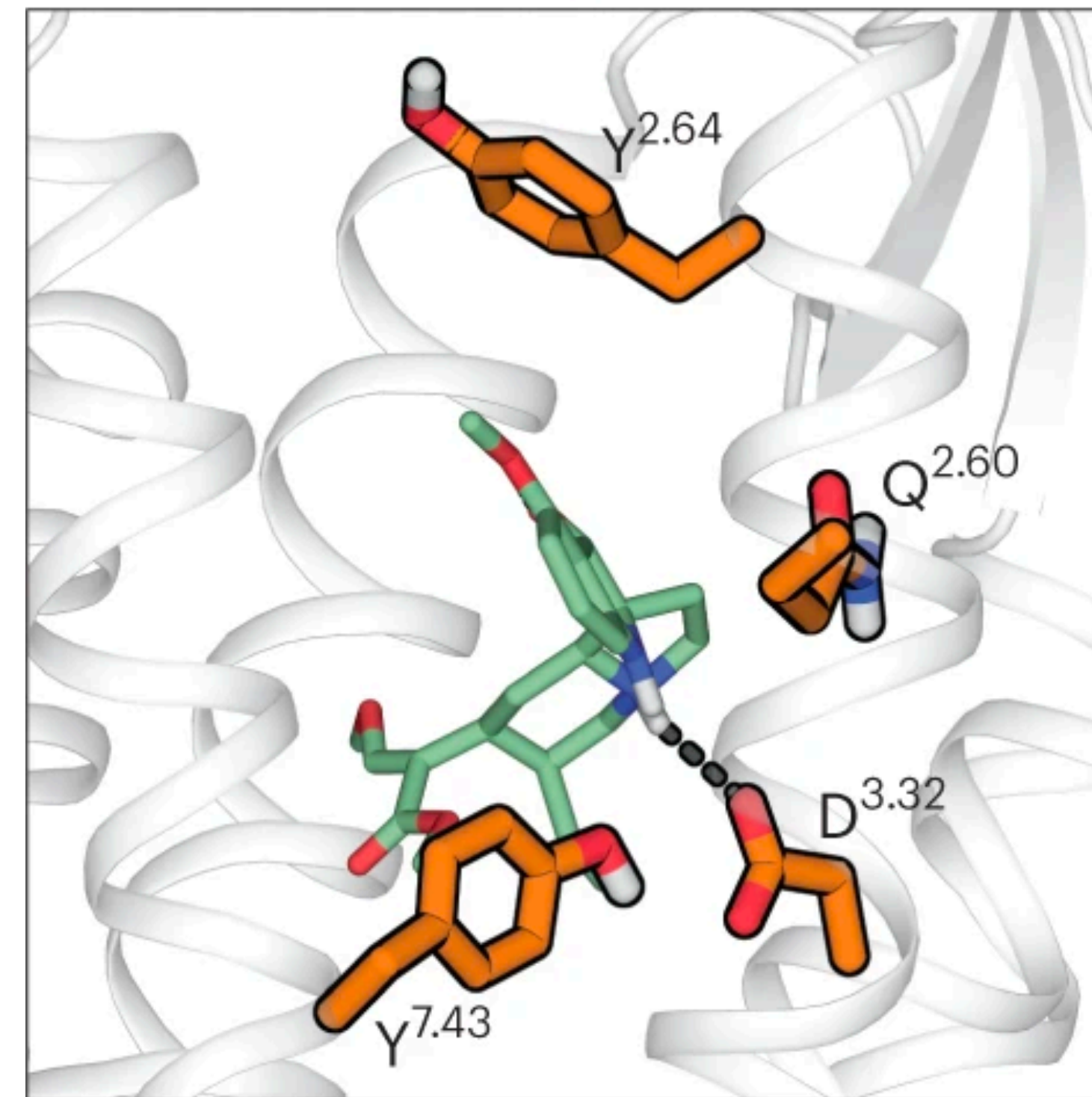


# Lofentaniol stabilizes a polar network holding Y7.43 inwards MP disrupts this polar network

Lofentaniol: Arrestin biased



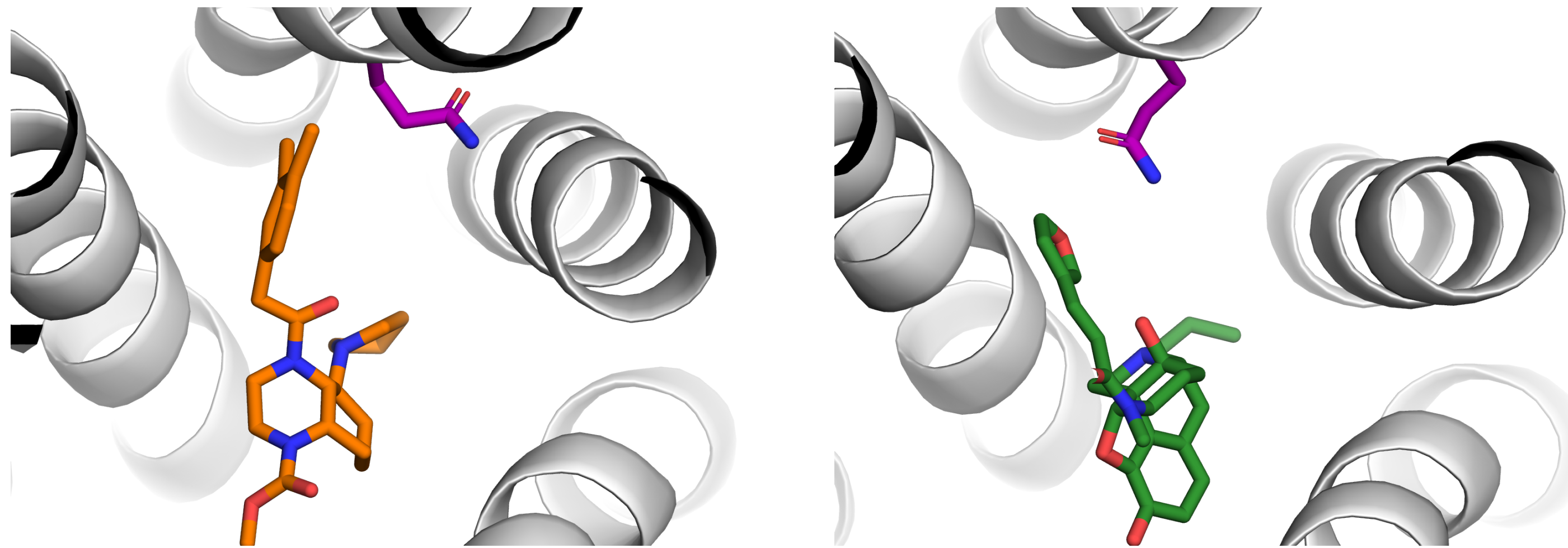
MP: G protein biased



Representative MD frames

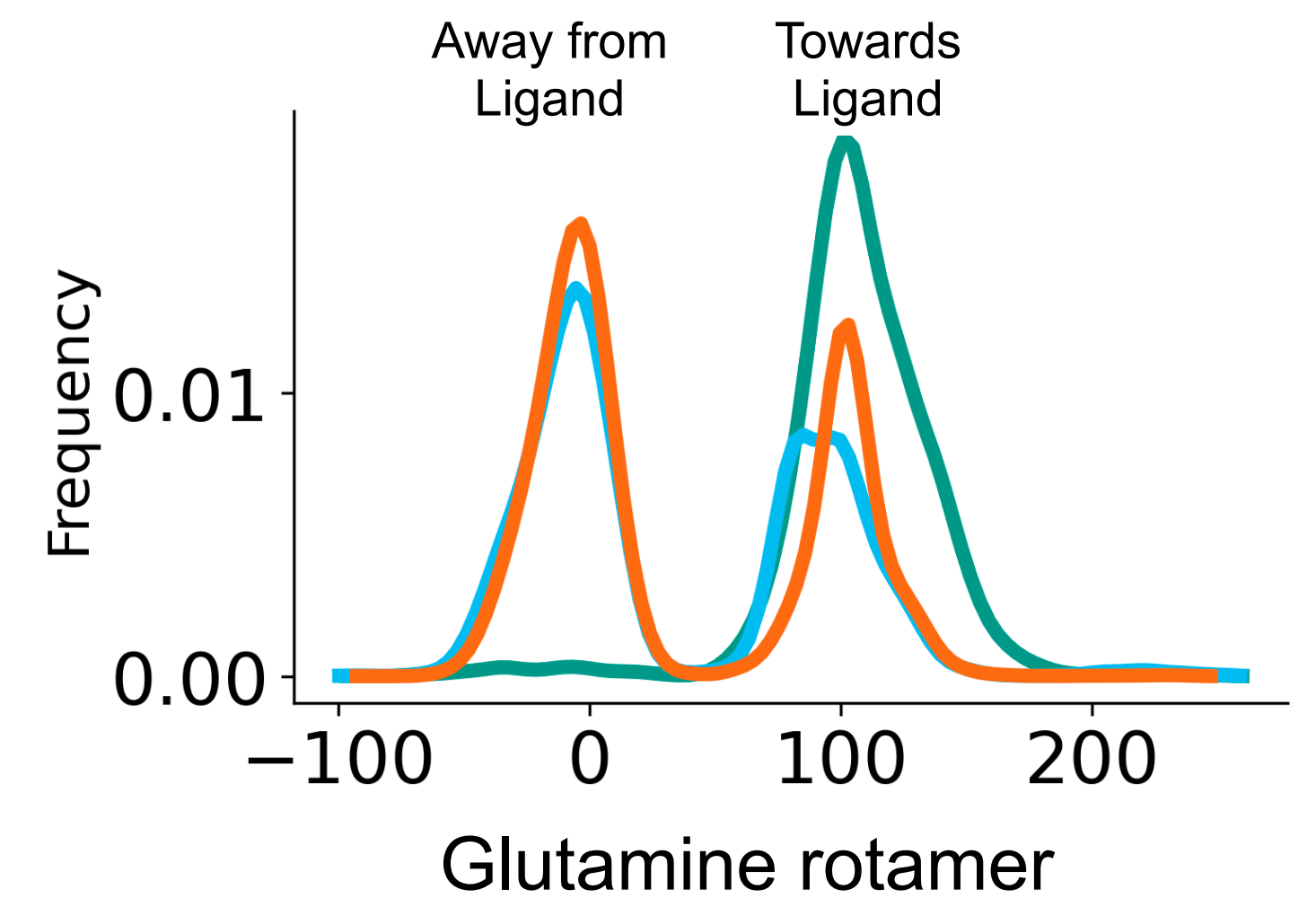
# Nalfurafine has a similar effect as MP, but through a different mechanism

MD frames

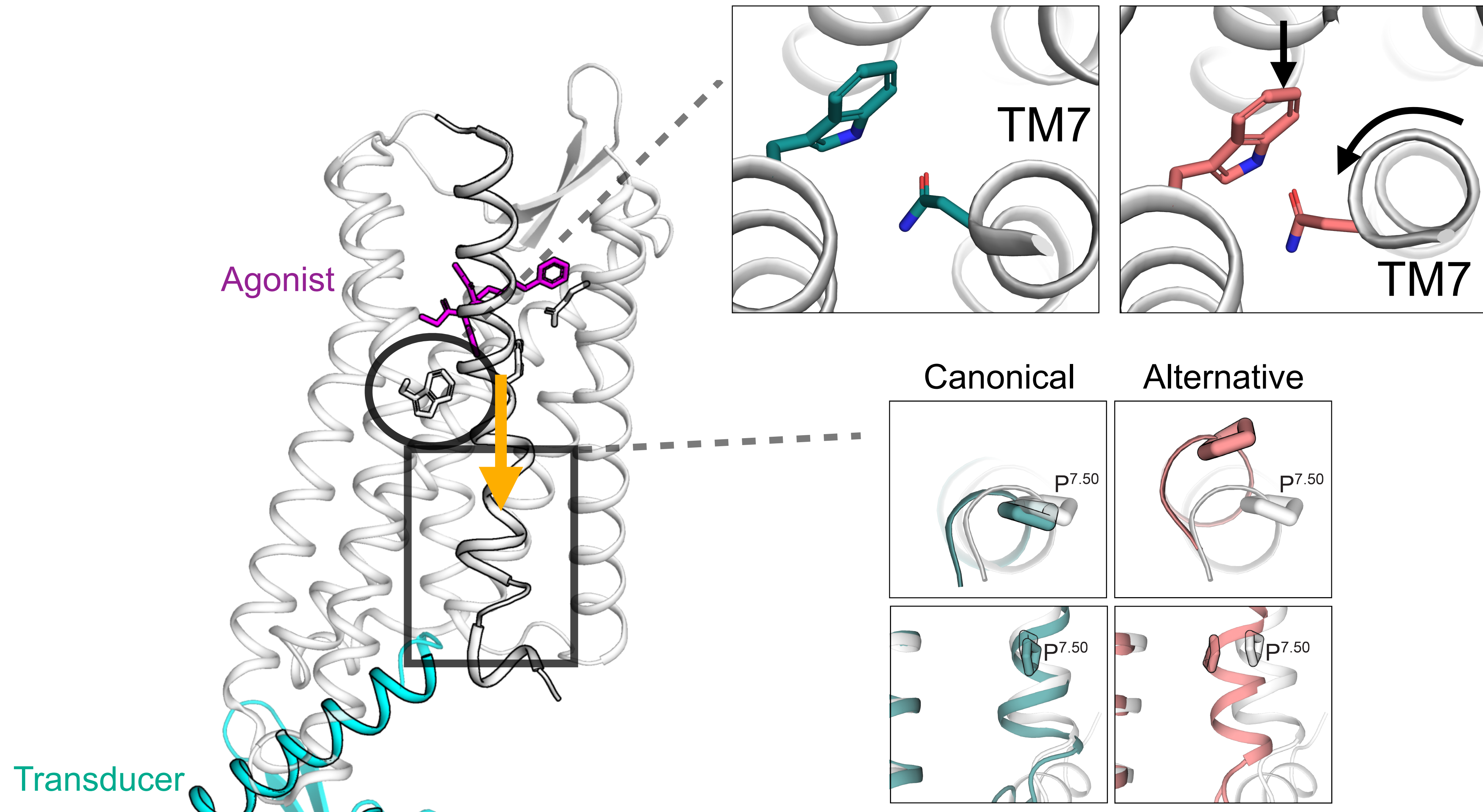


Extracellular view

Nalfurafine: G protein biased  
U50,488: Balanced  
WMS-X600: Arrestin biased



# Tryptophan pathway



# Vertical displacement of tryptophan explains arrestin-bias of WMS-X600

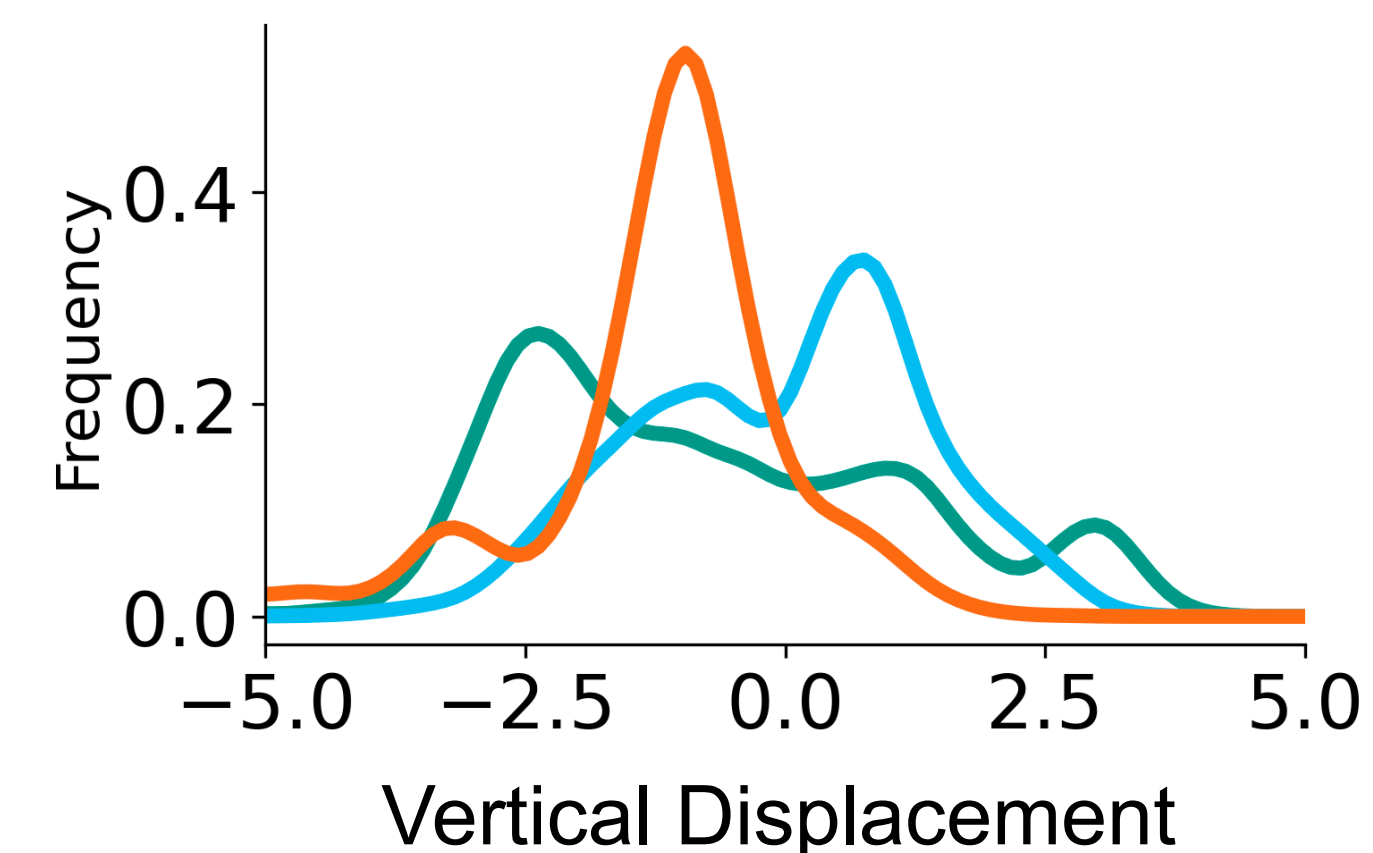
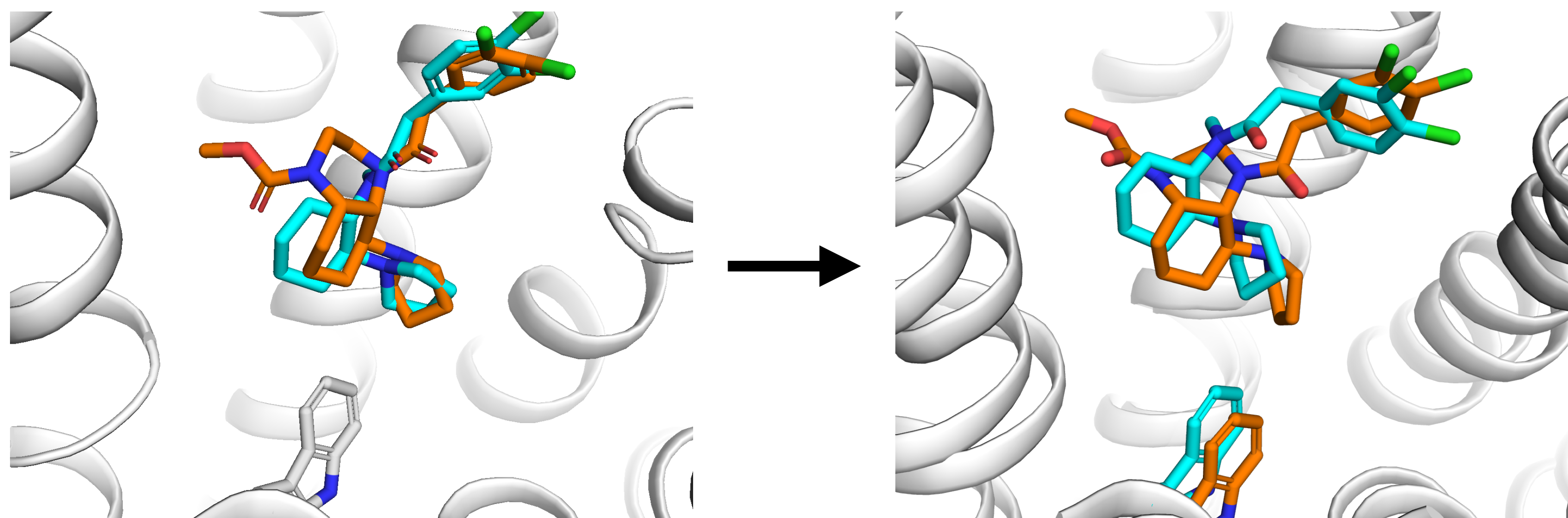
Nalfurafine: G protein biased

U50,488: Balanced

WMS-X600: Arrestin biased

Initial docked poses

MD frame



\*Nalfurafine not significantly different than others

- Mutating this tryptophan to an alanine removes bias between WMS-X600 and U50,488
- Nalfurafine becomes even more G protein biased. Opportunity!

# Acknowledgements

- Ron Dror
- Deniz Aydin
- Yianni Laloudakis
- Carl-Mikael Suomivuori
- Naomi Latorraca
- Stephan Eismann
- Matthew King
- Scott Hollingsworth
- **Qianhui Qu**
- **Weijiao Huang**
- **Alpay B. Seven**
- **Susruta Majumdar**
- **Brian K. Kobilka**
- **Georgios Skiniotis**
- Haoqing Wang
- Soumen Chakraborty
- Michael J. Robertson
- Asuka Inoue
- Bryan L. Roth
- **Amal El Daibani**
- **Kuglae Kim**
- **Tao Che**
- Petr Popov
- Sarah M. Bernhard
- Brian E. Krumm
- Reid H. J. Olsen
- Jeffrey F. DiBerto
- Ivy Carroll
- Vsevolod Katritch
- Bernhard Wunsch